



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(81) International Patent Classification ⁶ :	A2	(11) International Publication Number:	WO 96/34935
C11D 3/386 // C12N 9/54		(43) International Publication Date:	7 November 1996 (07.11.96)

(21) International Application Number:	PCT/EP96/01610	(81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).
(22) International Filing Date:	12 April 1996 (12.04.96)	
(30) Priority Data:		
95201161.7 5 May 1995 (05.05.95)	EP	
(34) Countries for which the regional or international application was filed:	NL et al.	
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(54) Title: SUBTILISIN VARIANTS

(57) Abstract

Detergent compositions are provided comprising variants of subtilase, which have been produced by mutating the genes for a number of subtilases and expressing the mutated genes in suitable hosts. The enzymes exhibit improved stability and/or wash performance in detergents, especially liquid detergents and soap bars, in comparison to their wild type parent enzymes. The enzymes are well-suited for use in liquid detergent compositions and soap bars.

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SUBTILISIN VARIANTS

TECHNICAL FIELD

This invention relates to novel mutant enzymes or 5 enzyme variants useful in formulating detergent compositions and exhibiting improved storage stability while retaining or improving their wash performance; cleaning and detergent compositions containing said enzymes; mutated genes coding for the expression of said enzymes when inserted into a 10 suitable host cell or organism; and such host cells transformed therewith and capable of expressing said enzyme variants.

15 BACKGROUND OF THE INVENTION

In the detergent industry enzymes have for more than 30 years been implemented in washing formulations. Enzymes used in such formulations comprise proteases, 20 lipases, amylases, cellulases, as well as other enzymes, or mixtures thereof. Commercially most important are proteases.

Although proteases have been used in the detergent industry for more than 30 years, much remains unknown as to details of how these enzymes interact with substrates and/other substances present in e.g. detergent compositions. 25 Some factors related to specific residues and influencing certain properties, such as oxidative and thermal stability in general have been elucidated, but much remains to be found out. Also, it is still not exactly known which physical or chemical characteristics are responsible for a 30 good washing performance or ability of a protease in a specific detergent composition.

The currently used proteases have for the most part been found by isolating proteases from nature and testing them in detergent formulations.

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PROTEASES

Enzymes cleaving the amide linkages in protein substrates are classified as proteases, or (interchangeably)

peptidases (see Walsh, 1979, *Enzymatic Reaction Mechanisms*, W.H. Freeman and Company, San Francisco, Chapter 3). Bacteria of the *Bacillus* species secrete two extracellular species of protease, a neutral, or metalloprotease, and an 5 alkaline protease which is functionally a serine endopeptidase and usually referred to as subtilisin. Secretion of these proteases has been linked to the bacterial growth cycle, with greatest expression of protease during the stationary phase, when sporulation also occurs. 10 Joliffe et al. (1980) *J. Bacteriol.* 141 1199-1208, have suggested that *Bacillus* proteases function in cell wall turnover.

SUBTILASES

15 A serine protease is an enzyme which catalyses the hydrolysis of peptide bonds, and in which there is an essential serine residue at the active site (White, Handler and Smith, 1973 "Principles of Biochemistry," Fifth Edition, McGraw-Hill Book Company, NY, pp. 271-272). 20 The bacterial serine proteases have molecular weights in the 20,000 to 45,000 range. They are inhibited by diisopropyl-fluorophosphate. They hydrolyse simple terminal esters and are similar in activity to eukaryotic chymotrypsin, also a serine protease. A more narrow term, 25 alkaline protease, covering a sub-group, reflects the high pH optimum of some of the serine proteases, from pH 9.0 to 11.0 (for review, see Priest (1977) *Bacteriological Rev.* 41 711-753).

30 A sub-group of the serine proteases tentatively designated subtilases has been proposed by Siezen et al., *Protein Engng.* 4 (1991) 719-737. They are defined by homology analysis of more than 40 amino acid sequences of serine proteases previously referred to as subtilisin-like proteases. A subtilisin was previously defined as a serine 35 protease produced by Gram-positive bacteria or fungi, and according to Siezen et al. now is a subgroup of the subtilases. A wide variety of subtilisins have been identified, and the amino acid sequence of a number of

subtilisins have been determined. These include more than six subtilisins from *Bacillus* strains, namely, subtilisin 168, subtilisin BPN', subtilisin Carlsberg, subtilisin Y, subtilisin amylosacchariticus, and mesenteropeptidase 5 (Kurihara et al. (1972) *J. Biol. Chem.* 247 5629-5631; Wells et al. (1983) *Nucleic Acids Res.* 11 7911-7925; Stahl and Ferrari (1984) *J. Bacteriol.* 159 811-819, Jacobs et al. (1985) *Nucl. Acids Res.* 13 8913-8926; Nedkov et al. (1985) *Biol. Chem. Hoppe-Seyler* 366 421-430, Svendsen et al. (1986) 10 *FEBS Lett.* 196 228-232), one subtilisin from an actinomycetales, thermitase from *Thermoactinomyces vulgaris* (Meloun et al. (1985) *FEBS Lett.* 198 195-200), and one fungal subtilisin, proteinase K from *Tritirachium album* (Jany and Mayer (1985) *Biol. Chem. Hoppe-Seyler* 366 584-492). 15 for further reference Table I from Siezen et al. has been reproduced below.

Subtilisins are well-characterized physically and chemically. In addition to knowledge of the primary structure (amino acid sequence) of these enzymes, over 50 high 20 resolution X-ray structures of subtilisins have been determined which delineate the binding of substrate, transition state, products, at least three different protease inhibitors, and define the structural consequences for natural variation (Kraut (1977) *Ann. Rev. Biochem.* 46 331-25 358).

In the context of this application substrate should be interpreted in its broadest form as comprising a compound containing at least one peptide bond susceptible to hydrolysis by a subtilisin protease.

30 Also the expression "product" should in the context of this invention be interpreted to include the products of a hydrolysis reaction involving a subtilisin protease. A product may be the substrate in a subsequent hydrolysis reaction.

35 One subgroup of the subtilases, I-S1, comprises the "classical" subtilisins, such as subtilisin 168, subtilisin BPN', subtilisin Carlsberg (ALCALASE®, NOVO NORDISK A/S), and subtilisin DY.

A further subgroup of the subtilases I-S2, is recognised by Siezen et al. (supra). Sub-group I-S2 proteases are described as highly alkaline subtilisins and comprise enzymes such as subtilisin PB92 (MAXACAL®, Gist-Brocades NV), subtilisin 309 (SAVINASE®, NOVO NORDISK A/S), subtilisin 147 (ESPERASE®, NOVO NORDISK A/S), and alkaline elastase YaB.

In the context of this invention, a subtilase variant or mutated subtilase means a subtilase that has been 10 produced by an organism which is expressing a mutant gene derived from a parent microorganism which possessed an original or parent gene and which produced a corresponding parent enzyme, the parent gene having been mutated in order to produce the mutant gene from which said mutated 15 subtilisin protease is produced when expressed in a suitable host.

Random and site-directed mutations of the subtilase gene have both arisen from knowledge of the physical and chemical properties of the enzyme and 20 contributed information relating to subtilase's catalytic activity, substrate specificity, tertiary structure, etc. (Wells et al. (1987) *Proc. Natl. Acad. Sci. U.S.A.* 84: 1219-1223; Wells et al. (1986) *Phil. Trans. R. Soc. Lond.A.* 317: 415-423; Hwang and Marshel (1987) *Biochem.* 26: 2669-2673; Rao 25 et al., (1987) *Nature* 328: 551-554.

More recent publications covering this area are Carter et al. (1989) *Proteins* 6: 240-248 relating to design of variants that cleave a specific target sequence in a substrate (positions 24 and 64); Graycar et al. (1992) 30 *Annals of the New York Academy of Sciences* 672: 71-79 discussing a number of previously published results; and Takagi (1993) *Int. J. Biochem.* 25: 307-312 also reviewing previous results.

Especially site-directed mutagenesis of the subtilisin genes has attracted much attention, and various mutations are described in the following patent applications and 35 patents:

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Reissue Patent No. 34,606 (GENENCOR) relating to site specific or randomly generated mutations in "carbonyl hydrolases" and subsequent screening of the mutated enzymes for various properties, such as K_{cat}/K_m ratio, pH-activity profile, and oxidation stability. This publication reveals that site-specific mutation is feasible, and that mutation of subtilisin BPN' in certain specified positions, i.e. 5 ^{10}Tyr , ^{12}Asp , ^{13}Asn , ^{14}Tyr , ^{22}Met , ^{16}Gly , ^{18}His , ^{19}Gly , ^{18}Phe , ^{21}Ser , ^{22}Ser , ^{23}Tyr , ^{15}Glu or ^{17}Ala , provide for enzymes 10 exhibiting altered properties. Since these positions all except position -1 were known to be involved in the functioning of the enzyme prior to the filing of the application, and therefore evident to select, this application does not contribute much to solving the problem 15 of deciding where to introduce mutations in order to obtain enzymes with desired properties.

EP-A-214 435 (HENKEL) relating to cloning and expression of subtilisin Carlsberg and two mutants thereof. In this application no reason for mutation of ^{15}Asp to ^{15}Ser 20 and ^{16}Ser to ^{16}Asp is provided.

In International patent publication WO-A-87/04461 (AMGEN) it is proposed to reduce the number of Asn-Gly sequences present in the parent enzyme in order to obtain mutated enzymes exhibiting improved pH and heat stabilities, 25 in the application emphasis is put on removing, mutating, or modifying the ^{10}Asn and the ^{21}Asn residues in subtilisin BPN'. No examples are provided for any deletions or for modifying the Gly-residues.

International patent publication WO-A-87/05050 30 (GENEX) discloses random mutation and subsequent screening of a large number of mutants of subtilisin BPN' for improved properties. In the application mutations are described in positions ^{11}Asn , ^{13}Gly , ^{24}Thr , ^{18}Gly , ^{16}Ala , ^{18}Ser , ^{12}Leu , and ^{23}Ser .

35 In EP-A-251 446 (GENENCOR) it is described how homology considerations at both primary and tertiary structural levels may be applied to identify equivalent amino acid residues whether conserved or not. This

information together with the inventors knowledge of the tertiary structure of subtilisin BPN' lead the inventors to select a number of positions susceptible to mutation with an expectation of obtaining mutants with altered properties.

5 The positions so identified are: ¹²²Met, ²²²Met, ¹⁰¹Tyr, ¹⁵²Ala, ¹⁵⁶Glu, ¹⁶⁶Gly, ¹⁶⁹Gly, ¹⁸⁹Phe, ²¹⁷Tyr. Also ¹⁸⁵Asn, ²¹Tyr, ²²Thr, ²⁴Ser, ²²Asp, ³³Ser, ³⁶Asp, ⁴⁶Gly, ⁵⁶Ala, ⁶⁶Ser, ⁷⁶Met, ⁷⁷Asn, ⁷⁷Ser, ⁹⁶Lys, ⁹⁸Val, ⁹⁶Leu, ¹⁰⁷Ile, ¹¹⁸Gly, ¹⁷⁰Lys, ¹⁷¹Tyr, ¹⁷²Pro, ¹⁷³Asp, ¹⁹³Met, ²⁰⁴Ser, ²¹⁰Lys, and ²²³Ser, which positions are
10 identified as being expected to influence various properties of the enzyme. Also, a number of mutations are exemplified to support these suggestions. In addition to single mutations in these positions the inventors also performed a number of multiple mutations. Further the inventors identify
15 ²¹⁵Gly, ⁶⁶His, ¹²⁶Leu, ¹³⁵Leu, and amino acid residues within the segments 97-103, 126-129, 213-215, and 152-172 as having interest, but mutations in any of these positions are not exemplified.

Especially of interest for the purpose of the
20 present invention the inventors of EP-A-251 446 suggest to substitute ¹⁷⁸Lys (in subtilisin BPN', type I-S1), specifically they suggest to introduce Glu or Arg for the original Lys. It appears that the Glu variant was produced and it was found that it was highly susceptible to autolytic
25 degradation (cf. pages 48, 121, 123 (Table XXI includes an obvious error, but indicates a reduction in autolysis halftime from 86 to 13 minutes) and Fig. 32).

EP-A-260 105 (GENENCOR) describes modification of certain properties in enzymes containing a catalytic triad
30 by selecting an amino acid residue within about 15 Å from the catalytic triad and replace the selected amino acid residue with another residue. Enzymes of the subtilase type described in the present specification are specifically mentioned as belonging to the class of enzymes containing a catalytic triad. In subtilisins positions 222 and 217 are indicated as preferred positions for replacement.

Also, it has been shown by Thomas, Russell, and Persht (1985) Nature 318 375-376 that exchange of ²²Asp into

"Ser in subtilisin BPN' changes the pH dependency of the enzyme.

In a subsequent article (1987) *J. Mol. Biol.* 193 803-813, the same authors also discuss the substitution of 5 "Ser in place of "Glu. Both these mutations are within a distance of about 15Å from the active "His.

In *Nature* 328 496-500 (1987) Russel and Persht discuss the results of their experiments and present rules 10 for changing pH-activity profiles by mutating an enzyme to obtain changes in surface charge.

WO-A-88/08028 (Genex) and WO-A-88/08033 (Amgen) both relate to modifications of amino acid residues in the calcium binding sites of subtilisin BPN'. The enzyme is said to be stabilized by substituting more negatively charged 15 residues for the original ones.

In WO-A-89/06279 (NOVO NORDISK A/S) position 170 is indicated as interesting and it is suggested to replace the existing residue with Tyr. However, no data are given in respect of such a variant. In WO-A-91/00345 (NOVO NORDISK 20 A/S) the same suggestion is made, and it is shown that the Tyr variant of position 170 in subtilisin 309 (type I-S2) exhibits an improved wash performance in detergents at a pH of about 8 (variant S003 in Tables III, IV, V, VI, VIII, X). The same substitution in combination with other 25 substitutions in other positions also indicates an improved wash performance (S004, S011-S014, S022-S024, S019, S020, S203, S225, S227 in the same table and Table VII) all in accordance with the generic concept of said application.

In EP-A-525 610 (SOLVAY) it is suggested to 30 improve the stability of the enzyme (a type I-S2 subtilase closely related to subtilisin PB92) towards ionic tensides by decreasing the hydrophobicity in certain surface regions thereof. It is consequently suggested to substitute Gln for the Arg in position 164 (170 if using BPN' numbering). No 35 variants comprising this substitution are disclosed in the application.

In WO-A-94/02618 (GIST-BROCADES N.V.) a number of position 164 (170 if using BPN' numbering) variants of the

I-S2 type subtilisin PB92 are described. Examples are provided showing substitution of Met, Val, Tyr, Ile, for the original Arg. Wash performance testing in powder detergents of the variants indicates a slight improvement.

5 Especially for the Ile variant wash performance tests on cacao an improvement of about 20-30% is indicated. No stability data are provided.

10 INDUSTRIAL APPLICATIONS OF SUBTILASES

Proteases such as subtilisins have found much utility in industry, particularly in detergent formulations, as they are useful for removing proteinaceous stains.

15 At present at least the following proteases are known to be commercially available and many of them are marketed in large quantities in many countries of the world:

Subtilisin BPN' or Novo, available from e.g. SIGMA, St. Louis, U.S.A.

20 Subtilisin Carlsberg, marketed by NOVO NORDISK A/S (Denmark) as ALCALASE® and by IBIS (Holland) as MAXATASE®; Both of these belong to subtilase subgroup I-S1.

Among the subtilase sub-group I-S2 the following are known to be marketed:

25 A *Bacillus latus* subtilisin, subtilisin 309, marketed by NOVO NORDISK A/S (Denmark) as SAVINASE®. A protein engineered variant of this enzyme is marketed as DURAZYM®.

30 Enzymes closely resembling SAVINASE®, such as subtilisin PB92, MAXACAL® marketed by Gist-Brocades N.V. (a protein engineered variant of this enzyme is marketed as MAXAPEM®), OPTICLEAN® marketed by SOLVAY et Cie. and PURAFECT® marketed by GENENCOR International.

A *Bacillus latus* subtilisin, subtilisin 147, marketed by NOVO NORDISK A/S (Denmark) as ESPERASE®;

35 To be effective, however, such enzymes must not only exhibit activity under washing conditions, but must also be compatible with other detergent components during detergent production and storage.

3

For example, subtilisins may be used in combination with other enzymes active against other substrates, and the selected subtilisin should possess stability towards such enzymes, and also the selected 5 subtilisin preferably should not catalyze degradation of the other enzymes. Also, the chosen subtilisin should be resistant to the action from other components in the detergent formulation, such as bleaching agents, oxidizing agents, etc., in particular an enzyme to be used in a detergent 10 formulation should be stable with respect to the oxidizing power, calcium binding properties, and pH conditions rendered by the non-enzymatic components in the detergent during storage and in the wash liquor during wash.

The ability of an enzyme to catalyze the 15 degradation of various naturally occurring substrates present on the objects to be cleaned during e.g. wash is often referred to as its washing ability, washability, detergency, or wash performance. Throughout this application the term wash performance will be used to encompass this 20 property.

The ability of an enzyme to remain active in the presence of other components of a detergent composition prior to being put to use (normally by adding water in the washing process) is usually referred to as storage stability 25 or shelf life. It is often measured as half-life, $t_{1/2}$. We will use the expression storage stability for this property throughout this application to encompass this property.

Naturally occurring subtilisins have been found to possess properties which are highly variable in relation to 30 their washing power or ability under variations in parameters such as pH. Several of the above marketed detergent proteases, indeed, have a better performance than those marketed about 20 years ago, but for optimal performance each enzyme has its own specific conditions regarding 35 formulation and wash conditions, e.g. pH, temperature, ionic strength (=I), active system (tensides, surfactants, bleaching agent, etc.), builders, etc.

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As a consequence it is found that an enzyme possessing desirable properties at low pH and low I may be less attractive at more alkaline conditions and high I, or an enzyme exhibiting fine properties at high pH and high I 5 may be less attractive at low pH, low I conditions.

Also, it has been found that the storage stability differs between the enzymes, but it has further been found that a specific enzyme exhibits large variations in storage stability in respect of different detergent formulations, 10 dependent upon a number of parameters, such as pH, pI, bleach system, tensides, etc., and upon the physical state of the detergent compositions, which may be in powder, dust, or liquid form. Furthermore it may be concentrated or dilute.

15 The advent and development of recombinant DNA techniques has had a profound influence in the field of protein chemistry.

Through the application of this technology it is possible now to construct enzymes having desired amino acid 20 sequences, and as indicated above a fair amount of research has been devoted to designing subtilisins with altered properties.

Among the proposals the technique of producing and screening a large number of mutated enzymes as described in 25 EP-A-130 756 (GENENTECH) (US Reissue Patent No. 34,606 (GENENCOR)) and International patent publ. no. WO-A-87/05050 (GENEX) correspond to a large extend to the classical method of isolating native enzymes, submit them to classical mutagenesis programs (using radiation or chemical mutagens) 30 and screen them for their properties. The difference lies in that these methods are more efficient through the knowledge of the presence of a large number of variant enzymes substituted in a specific position.

A subtilisin enzyme typically comprises about 275 35 amino acid residues. Each residue is capable of being 1 out of 20 possible naturally occurring amino acids. Therefore one very serious draw-back in that procedure is the very large number of mutations generated that have to be

submitted to a number of preliminary screenings to determine their properties.

A procedure as outlined in these patent applications will consequently only be slightly better than 5 the traditional random mutation procedures which have been known for years.

The other known techniques relate to changing specific properties, such as oxidation stability, thermal stability, Ca-stability, transesterification and hydrolysis 10 rate (EP-A-260 105 (GENENCOR)), pH-activity profile (Thomas, Russell, and Persht, *supra*), and substrate specificity (International patent publ. WO-A-88/07578 (GENENTECH)). None of these publications relates to changing either the wash performance of enzymes or their storage stability.

15 In International Patent Application no. PCT/DK 88/00002 (NOVO NORDISK A/S) it is proposed to use the concept of homology comparison to determine which amino acid positions should be selected for mutation and which amino acids should be substituted in these positions in order to 20 obtain a desired change in wash performance.

By using such a procedure the task of screening is reduced drastically, since the number of mutants generated is much smaller, but with that procedure it is only foreseen that enzymes exhibiting the combined useful properties of 25 the parent enzyme and the enzyme used in the comparison may be obtained.

Thus, as indicated above no relationship has yet been identified between well defined properties of an enzyme such as those mentioned above and the wash performance and 30 storage stability of an enzyme in various detergent compositions.

The problem seems to be that although much research has been directed at revealing the mechanism of enzyme activity, still only little is known about the 35 factors in structure and amino acid residue combination that determine the properties, such as storage stability in detergents, of enzymes in relation to most of their characteristics, especially when the enzymes are present in

complex mixtures.

Consequently, there still exists a need for further improvement and tailoring of enzymes to detergent systems, as well as a better understanding of the mechanism 5 of protease action and degradation in the practical use of cleaning or detergent compositions. Such an understanding could result in rules which may be applied for selecting mutations that with a reasonable degree of certainty will result in an enzyme exhibiting improved storage stability 10 and/or performance under specified conditions in a detergent composition.

SUMMARY OF THE INVENTION

It has now surprisingly been found that a 15 subtilase variant having improved storage stability and/or improved performance in detergent compositions can be obtained by substituting one or more amino acid residues situated in, or in the vicinity of a hydrophobic domain of the parent subtilase for an amino acid residue more 20 hydrophobic than the original residue, said hydrophobic domain comprising the residues corresponding to residues P129, P131, I165, Y167, Y171 of BLS309 (in BASBPN numbering), and said residues in the vicinity thereof comprises residues corresponding to the residues E136, G159, 25 S164, S164, R170, A194 and 195 of BLS309 (in BASBPN numbering), with the exception of the R170M, R170I and R170V variants of BABP92.

The present invention relates consequently in its first aspect to enzyme variants exhibiting improved 30 stability and/or wash performance in detergents, in particular in liquid detergents, especially concentrated liquid detergents, and in soap bars.

In its second aspect the invention relates to DNA constructs capable of expressing the enzymes of the first 35 aspect, when inserted in a suitable manner into a host cell that subsequently is brought to express the subtilisin enzyme(s) of the first aspect.

In a third aspect the invention relates to the

production of the subtilisin enzymes of the invention by inserting a DNA construct according to the second aspect into a suitable host, cultivating the host to express the desired subtilase enzyme, and recovering the enzyme product.

5 The invention relates, in part, but is not limited to, mutants of the genes expressing the subtilase sub-group I-S2 enzymes and the ensuing enzyme variants, as indicated above.

10 Other subtilase gene variants encompassed by the invention are such as those of the subtilase subgroup I-S1, e.g. Subtilisin BPN', and Subtilisin Carlsberg genes and ensuing variant Subtilisin BPN', Proteinase K, and Subtilisin Carlsberg enzymes, which exhibit improved stability and/or wash performance in detergents.

15 Still further subtilase gene variants encompassed by the invention are such as Proteinase K and other genes and ensuing variant Proteinase K, and other subtilase enzymes, which exhibit improved stability and/or performance in detergents.

20 Other examples of parent subtilase enzymes that can be modified in accordance with the invention are listed in Table I.

25 Further the invention relates to the use of the mutant enzymes in cleaning compositions and cleaning compositions comprising the mutant enzymes, especially detergent compositions comprising the mutant subtilisin enzymes. Specifically the invention relates to liquid detergent compositions, especially concentrated liquid detergents and to soap bars comprising such enzyme variants.

ABBREVIATIONSAMINO ACIDS

A	=	Ala	=	Alanine	
V	=	Val	=	Valine	
S	L	=	Leu	=	Leucine
I	=	Ile	=	Isoleucine	
P	=	Pro	=	Proline	
F	=	Phe	=	Phenylalanine	
W	=	Trp	=	Tryptophan	
10	M	=	Met	=	Methionine
G	=	Gly	=	Glycine	
S	=	Ser	=	Serine	
T	=	Thr	=	Threonine	
C	=	Cys	=	Cysteine	
15	Y	=	Tyr	=	Tyrosine
N	=	Asn	=	Asparagine	
Q	=	Gln	=	Glutamine	
D	=	Asp	=	Aspartic Acid	
E	=	Glu	=	Glutamic Acid	
20	K	=	Lys	=	Lysine
R	=	Arg	=	Arginine	
H	=	His	=	Histidine	

NUCLEIC ACID BASES

25	A	=	Adenine
	G	=	Guanine
	C	=	Cytosine
	T	=	Thymine (only in DNA)
	U	=	Uracil (only in RNA)

30

VARIANTS

In describing the various enzyme variants produced or contemplated according to the invention, the following nomenclatures have been adapted for ease of reference:

35 Original amino acid(s) - position(s) - substituted amino acid(s).

According to this the substitution of Glutamic acid for glycine in position 195 is designated as:

15

Gly 195 Glu or G195E

a deletion of glycine in the same position is:

Gly 195 * or G195*

and insertion of an additional amino acid residue such as

5 lysine is:

Gly 195 GlyLys or G195GK

Where a deletion is indicated in Table I, or
present in a subtilisin not indicated in Table I, an
10 insertion in such a position is indicated as:

* 36 Asp or *36D

for insertion of an aspartic acid in position 36.

Multiple mutations are separated by pluses, i.e.:

15 Arg 170 Tyr + Gly 195 Glu or R170Y+G195E
representing mutations in positions 170 and 195 substituting
tyrosine and glutamic acid for arginine and glycine, respectively.

20 POSITIONS

In describing the variants in this application and
in the appended claims use is made of the alignment of
various subtilases in Siezen et al., Supra. In other
publications relating to subtilases other alignments or the
25 numbering of specific enzymes have been used. It is a
routine matter for the skilled person to establish the
position of a specific residue in the numbering used here.
Reference is also made to Fig. 1 showing an alignment of
residues relevant for the present invention from a large
30 number of subtilases. Reference is also made to Table I of
WO-A-91/00345 showing an alignment of residues relevant for
the present invention from a large number of subtilases.

TABLE I

Presently established Subtilases (from Siezen et al., *supra*)

	Organism	cDNA, gene	enzyme	acronym
5	<u>PROKARYOTES</u>			
	<u>Bacteria: Gram-positive</u>			
	<i>Bacillus subtilis</i> 168	apr A	subtilisin 1168, spr	ABSS168
	<i>Bacillus amyloliquefaciens</i>		spr	
	subtilisin BPN' (NOVO)	BASBEN		
10	<i>Bacillus subtilis</i> DY	-	subtilisin DY	BSSDY
	<i>Bacillus licheniformis</i>	+	subtilisin Carlsberg	BLSCAR
	<i>Bacillus lentus</i>	+	subtilisin 147	BLS147
	<i>Bacillus alcalophilus</i> PB92		+	
	subtilisin PB92	BAPB92		
15	<i>Bacillus</i> sp. DSM 4828	-	alkaline protease	BDSM48
	<i>Bacillus</i> Yab	ale	alkaline elastase Yab	BYSYAB
	<i>Bacillus subtilis</i> 168	spr	min. extracell. prot.	BSESPR
	<i>Bacillus subtilis</i>	bpf	bacillopeptidase F	BSSPF
	<i>Bacillus subtilis</i> IFO3013	ispl	intracell.ser. prot. I	BSISPI
20	<i>Bacillus subtilis</i> AS0	-	intracell.ser. prot.	BSIAS0
	<i>Bacillus thuringiensis</i>	-	extracell. ser. prot.	BTTFINI
	<i>Bacillus cereus</i>	-	extracell. ser. prot.	BCESPR
	<i>Nocardiopsis dassonvillei</i>	-	alkaline ser. prot.	NDAPII
	<i>Thermoactinomyces vulgaris</i>	-	thermitase	TVTHER
25	<i>Enterococcus faecalis</i>	cyla	cytolysin component A	EFCYLA
	<i>Staphylococcus epidermidis</i>	epip	epidermin lead. prot.	SSEPIP
	<i>Streptococcus pyogenes</i>	scpA	CSa peptidase	SPSCPA
	<i>Lactococcus lactis</i> SK11	prtP	SK11 cell wall prot.	LLSK11
	<u>Bacteria: Gram-negative</u>			
30	<i>Dichelobacter nodosus</i>	+	basic protease	DNEBSPR
	<i>Xanthomonas campestris</i>	+	extracellular prot.	XCEXPFR
	<i>Serratia marcescens</i>	+	extracell. ser. prot.	SMEEXP
	<i>Thermus aquaticus</i> YT-1	pstI	aqualysin I	TRAQUA
	<i>Thermus</i> rT41A	+	T41A protease	TRT41A
35	<i>Vibrio alginolyticus</i>	proA	protease A	VAPROA
	<i>Streptomyces rutgersensis</i>	-	proteinase D	SRESFD
	<u>Archaea</u>			
	halophilic strain 172P1	-	halophil extra. prot.	ARB172
	(Table I, continued)			
40	Organism	cDNA, gene	enzyme	acronym
	<u>Cyanobacteria</u>			
	<i>Anabaena variabilis</i>	prch	Ca-dependent protease	AVPRCA
	<u>LOWER EUKARYOTES</u>			
	<u>Fungi</u>			
45	<i>Tritirachium album</i> Limber	+	proteinase K	TAPROK
	<i>Tritirachium album</i>	+	proteinase R	TAPROR

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	Triticum album	prot	proteinase T	TAPROT
	Aspergillus oryzae	+	alkaline protease	AOALPR
	Malbranchea pulchella	-	thermomycolin	MPTHMY
	Acremonium chrysogenum	alp	alkaline protease	ACALPR
5	<u>Yeasts</u>			
	Kluyveromyces lactis	kex1	Kex1 ser. proteinase	KLKEX1
	Saccharomyces cerevisiae	kex2	Kex2 ser. proteinase	SCKEX2
	Saccharomyces cerevisiae	prb1	protease B	SCPRL1
	Yarrowia lipolytica	xpr2	alk. extracell. prot.	YILXPR2
10				
	<u>HIGHER EUKARYOTES</u>			
	<u>Humans</u>			
	Caenorhabditis elegans	bli4	cuticle protease	CREBL4
	<u>Insects</u>			
15	Drosophila (fruit fly)	fur1	furin 1	DMFUR1
	Drosophila (fruit fly)	fur2	furin 2	DMFUR2
	<u>Plants</u>			
	Cucumis melo (melon)	-	cucumisin	CMCUCU
	<u>Mammals</u>			
20	Human (also rat, mouse)	fur	furin	HSFUR1
	Human (also mouse)	+	insulinoma PC2 prot.	HSIPC2
	Mouse	+	pituitary PC3 prot.	MMPPC3
	Human	+	tripeptidyl peptid. II	HSTPP

25

References used for Table I

References to amino acid sequences (GenBank®/EMBL Data Bank accession numbers are shown in brackets):

ARB172 Kamekura and Seno, (1990) *Biochem. Cell Biol.* 68 352-359 (amino acid sequencing of mature protease residues 1-35; residue I4 not determined).

30 BSS168 Stahl. and Ferrari. (1984) *J. Bacteriol.* 158, 411-418 (K01988). Yoshimoto, Oyama et al. (I488) *J. Biochem.* 103, 1060-1065 (the mature subtilisin from *B. subtilis* var. *amylosacchariticus* differs in having T130S and T162S). Svendsen, et al. (1986) *FEBS Lett.* 196, 228-232 (PIR A23624; amino acid sequencing; the mature alkaline mesentericopeptidase from *B. mesentericus* differs in having S85A, A88S, S89A, S183A and N259S).

35 40 BASBPN Wells, et al. (1983) *Nucl. Acids Res.* 11 7911-7925 (X00165). Vasantha et al., (1984) *J. Bacteriol.* 159 811-814 (K02496).

BSSDY Nedkov et al. (1983) *Hoppe-Seyler's Z. Physiol. Chem.* 364 1537-1540 (PIR A00969; amino acid sequencing).

5 BLSCAR Jacobs et al. (1985) *Nucleic Acids Res.* 13 8913-8926 (X03341). Smith et al. (1968) *J. Biol. Chem.* 243 2184-2191 (PIR A00968; amino acid sequencing; mature protease sequence differs in having T103S, P129A, S158W, N161S and S212N).

10 BLS147 Hastrup et al. (1989) PCT Patent Appl. WO 8906279. Pub. July 13 1989. (Esperase[®] from *B. latus*). Takami et al. (1990) *Appl. Microbiol. Biotechnol.*, 33 519-523 (amino acid sequencing of mature alkaline protease residues 1-20 from *Bacillus* sp. no. AH-101; this sequence differs from BLS147 in having N11S).

15 BABP92 van der Laan et al. (1991) *Appl. Environ. Microbiol.* 57 901-909. (Maxacal[®]). Hastrup et al. (1989) PCT Patent Appl. WO 8906279. Pub. 13 Jul 1989. (subtilisin 309. Savinase[®], from *B. latus* differs only in having N87S). Godette et al. (1991) Abstracts 5th Protein Society Symposium, June 6, Baltimore: abstract M8 (a high-alkaline protease from *B. latus* differs in having N87S, S99D, S101R, S103A, V104I and G159S).

20 25 BDSM48 Rettenmaier et al. (1990) PCT Patent Appl. WO 90/04022. Publ. April 19, 1990.

25 BYSYAB Kaneko et al. (1989) *J. Bacteriol.* 171 5232-5236 (M28537).

30 BSEPR Sloma et al. (1988) *J. Bacteriol.* 170 5557-5563 (M22407). Bruckner (1990) *Mol. Gen. Genet.* 221 486-490 (X53307).

35 BSBPF Sloma et al. (1990) *J. Bacteriol.* 172 1470-1477 (M29035; corrected). Wu et al. (1990) *J. Biol. Chem.* 265 6845-6850 (J05400; this sequence differs in having A169V and 586 less C-terminal residues due to a frameshift).

40 BSISP1 Koide et al. (1986) *J. Bacteriol.* 167 110-116 (M13760).

BSIA50 Strongin et al. (1978) *J. Bacteriol.* 133 1401-1411 (amino acid sequencing of mature protease residues 1-54; residues 3, 39, 40, 45, 46, 49 and 50 not determined).

5 BTFINI Chestukhina et al. (1985) *Biokhimiya* 50 1724-1730 (amino acid sequencing of mature protease residues 1-14 from *B. thuringiensis* variety *israeliensis*, and residues 1-16 and 223-243 from variety *finitimus*). Kunitate et al. (1989) *Agric. Biol. Chem.* 53 3251-3256 (amino acid sequencing of mature protease residues 6-20 from variety *kurstaki*. BTKURS).

10 BCESPR Chestukhina et al. (1985) *Biokhimiya* 50 1724-1730 (amino acid sequencing of mature residues 1-16 and 223-243).

15 NDAPII Tsujiibo et al. (1990) *Agric. Biol. Chem.* 54 2177-2179 (amino acid sequencing of mature residues 1-26).

20 TVTHER Meloun et al. (1985) *FEBS Lett.* 183 195-200 (PIR A00973; amino acid sequencing of mature protease residues 1-274).

25 RFCYLA Segarra et al. (1991) *Infect. Immun.* 59 1239-1246.

SEEPIP Schnell et al. (1991) personal communication (Siezen et al. (supra)).

SPSCPA Chen et al. (1990) *J. Biol. Chem.* 265 3161-3167 (J05224).

DNEBPR Kortt et al. (1991) Abstracts 5th Protein Society Symposium, June 22-26, Baltimore abstract S76.

30 LLSKII Vos et al. (1989) *J. Biol. Chem.* 264 13579-13585 (J04962). Kok et al. (1988) *Appl. Environ. Microbiol.* 54 231-238 (M24767; the sequence from strain Wg2 differs in 44 positions, including 18 differences in the protease domain, and a deletion of residues 1617-1676). Kiwaki et al. (1989) *Mol. Microbiol.* 3 359-369 (X14130; the sequence from strain NCD0763 differs in 46 positions, including 22 in the protease domain, and a deletion of residues 1617-1676).

20

XCEXPR Liu et al. (1990) *Mol. Gen. Genet.* 220 433-440.

SMEXSP Yanagida et al. (1986) *J. Bacteriol.* 166 937-934 (M13469).

5 TAAQUA Terada et al. (1990) *J. Biol. Chem.* 265 6576-6581 (J05414).

TRT41A McHale et al. (1990) Abstracts 5th Eur. Congr. Biotechn. Christiansen, Munck and Villadsen (eds), Munksgaard Int. Publishers, Copenhagen.

VAPROA Deane et al. (1989) *Gene* 76 281-288 (M25499).

10 SRESPD Lavrenova et al. (1984) *Biochemistry USSR.* 49 447-454 (amino acid sequencing of residues 1-23; residues 13, 18 and 19 not determined).

AVFRCA Maldener et al (1991) *Mol. Gen. Genet.* 225 113-120 (the published sequence has 28 uncertain residues 15 near position 200-210 due to a frameshift reading error).

TAPROK Gunkel and Gassen (1989) *Eur. J. Biochem.* 179 185-194 (X14688/XI4689). Jany et al. (1986) *J. Biol. Chem. Hoppe-Seyler* 267 87 (PIR A24541; amino acid sequencing; mature protease differs in having S745G, S11ST204-208DSL and VNL264-267FNL).

20 TAPROR Samal et al. (1990) *Mol. Microbiol.* 4 1789-1792 (X56116).

TAPROT Samal et al. (1989) *Gene* 85 329-333.

25 AOALPR Tatsumi et al. (1989) *Mol. Gen. Genet.* 219 33-38. Cheevadhanarah et al. (1991) EMBL Data Library (X54726).

MPTHMY Gaucher and Stevenson (1976) *Methods Enzymol.* 45 41S-433 (amino acid sequencing of residues 1-30 28, and hexapeptide LSGTGM with active site serine).

ACALPR Isogai et al. (1991) *Agric. Biol. Chem.* 55 471-477. Stepanov et al. (1986) *Int. J. Biochem.* 18 369-375 (amino acid sequencing of residues 1-27; the mature protease differs in having H13[1]Q, R13[2]N and S13[6]A).

35 KLXEX1 Tanguy-Rougeau, Wesolowski-Louvel and Fukuhara (1988) *FEBS lett.* 234 464-470 (X07038).

2)

SCREX2 Mizuno et al. (1988) *Biochem. Biophys. Res. Commun.* 156 246-254 (M24201).

SCPRB1 Moehle et al. (1987) *Mol. Cell. Biol.* 7 4390-4399 (M18097).

5 YLXYPR2 Davidow et al. (1987) *J. Bacteriol.* 169 4621-4629 (M17741). Matoba et al. (1988) *Mol. Cell Biol.* 8 4904-4916 (M23353).

CEBL14 Peters and Rose (1991) *The Worm Breeder's Gazette* 11 28.

10 DMFUR1 Roebroek et al. (1991) *FEBS Lett.* 289 133-137 (X59384).

DMFUR2 Roebroek et al. (1992) 267 17208-17215.

CMCUCU Kaneda et al. (1984) *J. Biochem.* 95 825-829 (amino acid sequencing of octapeptide NIISGTSM with active site serine).

15 HSFURI van den Ouwehand et al. (1990) *Nucl. Acids Res.* 18 664 (X04329) (the sequence of mouse furin differs in 51 positions, including five in the catalytic domain: A15E, Y21F, S223F, A232V and N258[2]D).

20 Misumi et al. (1990) *Nucl. Acids Res.* 18 6719 (X55660: the sequence of rat furin differs in 49 positions, including three in the catalytic domain: A15E, Y21F, H24R).

HSIPC2 Smeekens and Steiner (1990) *J. Biol. Chem.* 265 2997-3000 (J05252). Seidah et al. (1990) *DNA Cell Biol.* 9 415-424 (the sequence of mouse pituitary PC2 protease differs in 23 positions, including seven in the protease domain: I4F, S42[2]Y, E45D, N76S, D133E, V134L and G239[1]D).

30 MMPPC3 Smeekens et al. (1991) *Proc. Natl. Acad. Sci. USA* 88 340-344 (M58507). Seidah et al. (1990) *DNA Cell Biol.* 9 415-424 (M55668/M55669; partial sequence).

HSTPP Tomkinson and Jonsson (1991) *Biochemistry* 30 168-174 (J05299).

BRIEF DESCRIPTION OF THE FIGURES

In the drawings, Fig. 1 shows an alignment of a number of the subtilases mentioned in Table I;

5 Fig. 2 is a 3-dimensional representation of subtilisin 309 showing the location of the hydrophobic domain and some the amino acid residues in the vicinity thereof to be substituted according to the invention.

10 DETAILED DESCRIPTION OF THE INVENTION

It has surprisingly been found that the storage stability and/or performance of subtilases in detergents is improved when amino acid residues situated in the vicinity of a hydrophobic domain comprising the residues P129, P131, 15 I165, Y167, Y171 of subtilisin 309 are substituted for a more hydrophobic residue. The residues in question are especially R136, G159, S164, R170, A194, and G195.

Fig. 2 shows the hydrophobic domain in subtilisin 309 and residues in the vicinity thereof a number of which 20 are to be substituted in order to increase the hydrophobicity of the domain. This may be achieved by substituting hydrophobic residues for non-hydrophobic residues and/or by substituting residues to become even more hydrophobic than in the parent enzyme.

25 The same principle applies to the corresponding hydrophilic domain in other subtilases, the identification of which is within the skills of the average person working in this technical field. Graphic representations like the one in Fig. 2 can be produced for other subtilases to 30 determine the target residues to be substituted according to the invention.

A number hereof is indicated in Table II below:

Table II
residues in hydrophobic domain and in the vicinity thereof

Pos\Enz.	BASBPN	BLSCAR	BLS309	BLS147	TVTHER
5 domain					
129	P	A	P	T	T
131	G	G	P	G	G
165	V	I	I	V	P
167	Y	Y	Y	Y	Y
10 171	Y	Y	Y	Y	Y
15 Vicinity					
136	K	K	E	E	Q
159	S	S	G	G	T
164	T	T	S	G	A
170	K	K	R	R	Y
194	P	A	A	P	S
195	E	E	G	E	V

Table II was constructed using the alignment shown in Fig. 2. It is obvious that similar or larger tables covering other subtilases may easily be produced by the skilled person.

Consequently, the invention relates to subtilase variants in which the amino acid sequence has been changed through mutating the gene of the subtilisin enzyme, which it is desired to modify (the parent enzyme or gene), in the codon responsible for the expression of the amino acid residue in positions 129, 131, 165, 167, 171, 136, 159, 164, 170, 194, and 195, which residues are more hydrophobic than the residue(s) in the parent enzyme, especially such hydrophobic residues that comprise a relatively long

hydrophobic chain, such as Ile, Leu, and Val, whereby, when the mutated gene is expressed, the amino acid residue is substituted by a more hydrophobic residue, which increases the hydrophobicity of the domain as such.

Hydrophobic amino acid residues are generally the following: Val (V), Ile (I), Leu (L), Met (M), Phe (F), Pro (P) and Trp (W). Among these Val, Ile and Leu are preferred. By looking at Table II and applying the principle of the invention a number of candidates for substitution become clear.

For both BASBPN and BLSCAR it seems appropriate to make substitutions in positions 136, 159, 164, 167, 170 and 195. In BLS309 positions 136, 164, 167 and 170 would be the first choices, and positions 159 and 195 also would be a second choice. In BLS147 positions 136, 167, 170, and 195 are the first choice, while positions 159 and 164 are second. Finally, in TVTHER positions 136, 167 and 194 are the first choices, with 164 as a second one.

According to the invention it would entail an advantage to substitute the Gly residues in the hydrophobic domain to bulkier and more hydrophobic residues.

Such considerations apply for any hydrophilic or hydrophobic residue that may occupy any of the above mentioned position, meaning that any increase in hydrophobicity seems to be advantageous. This means that e.g. a very hydrophilic residue such as the charged residues Arg (R), Asp (D), Glu (E) or Lys (K) may be substituted by any residue that is less hydrophilic. Such less hydrophilic residues comprises the residues Gly (G), Cys (C), Ser (S), Ala (A), Thr (T), Tyr (Y), Gln (Q), His (H) or Asn (N).

Similar considerations can be applied to other subtilases having a hydrophobic domain in this part of the surface of the enzyme.

In the context of this invention a subtilase is defined in accordance with Siezen et al. *supra*. In a more narrow sense, applicable to many embodiments of the invention, the subtilases of interest are those belonging to the subgroups I-S1 and I-S2. In a more specific sense, many

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of the embodiments of the invention relate to serine proteases of gram-positive bacteria which can be brought into substantially unambiguous homology in their primary structure, with the subtilases listed in Table I above.

5 The present invention also comprises any one or more substitutions in the above mentioned positions in combination with any other substitution, deletion or addition to the amino acid sequence of the parent enzyme. Especially combinations with other substitutions known to 10 provide improved properties to the enzyme are envisaged.

Such combinations comprise the positions: 222 (improve oxidation stability), 218 (improves thermal stability), substitutions in the Ca-binding sites stabilising the enzyme, e.g. position 76, and many other 15 apparent from the prior art. Furthermore, combinations with the variants mentioned in EP-A-405 901 are also contemplated specifically.

VARIANTS

20 A: Single variants:

Subtilisin BPN', Subtilisin Carlsberg, Subtilisin 168, and Subtilisin DY variants:

25 K136V, K136I, K136L, K136M, K136F,
S159V, S159I, S159L, S159M, S159F,
T164V, T164I, T164L, T164M, T164F,
K170V, K170I, K170L, K170M, K170F,
E195V, E195I, E195L, E195M, E195F,

Thermitase variants:

30 Q136V, Q136I, Q136L, Q136M, Q136F,
T159V, T159I, T159L, T159M, T159F,
A164V, A164I, A164L, A164M, A164F,
Y167V, Y167I, Y167L, Y167M, Y167F,
Y170V, Y170I, Y170L, Y170M, Y170F,
35 S194V, S194I, S194L, S194M, S194F,

Subtilisin 309, Subtilisin 147, and Bacillus PB92 protease variants:

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E136V, E136I, E136L, E136M, E136F,
G159V, G159I, G159L, G159M, G159F,
G164V, G164I, G164L, G164M, G164F, (BLS147)
S164V, S164I, S164L, S164M, S164F, (BLS309 AND BAPB92)
5 Y167V, Y167I, Y167L, Y167M, Y167F,
R170V, R170I (both disclaimed for PAPB92),
R170L, R170M (disclaimed for PAPB92), R170F, R170G,
R170C,
A194V, A194I, A194L, A194M, A194F, (BLS309 AND BAPB92)
10 P194V, P194I, P194L, P194M, P194F, (BLS147)
E195V, E195I, E195L, E195M, E195F, (BLS147)
G195V, G195I, G195L, G195M, G195F, (BLS309 AND BAPB92)

15 B: Combination Variants:

Any of the above variants are contemplated to prove advantageous if combined with other variants in any of the positions: 27, 36, 57, 76, 101, 104, 123, 218, 222, 224
20 and 274.

Specifically the following BLS309 variants are considered appropriate for combination:

K27R, *36D, S57P, N76D, S101G, V104A, V104N, V104Y, N123S,
A194P, Q206E, N218S, M222S, M222A, T224S and T274A.

25 Also such variants comprising any one or two of the substitutions X167L, X167I, X170L, and/or X170I in combination with any one or more of the other substitutions, deletions and/or insertions mentioned above are advantageous.

30 Furthermore, variants comprising any of the variants V104N+S101G, K27R+V104Y+N123S+T274A, or N76D+V104A, in combination with any one or more of the substitutions, deletions and/or insertions mentioned above are deemed to exhibit improved properties.

35 Specific combinations to be mentioned are:

- a) S57P+R170L
- a') S57P+R170I
- b) R170L+N218S

- b') R170I+N218S
- c) S57P+R170L+N218S
- c') S57P+R170I+N218S
- c'') S57P+V104Y+R170L+N218S
- 5 c''') S57P+V104Y+R170I+N218S
- d) R170L+N218S+M222A
- d') R170I+N218S+M222S
- d'') R170L+N218S+M222A
- d'') R170I+N218S+M222S
- 10 e) S57P+R170L+S188P+A194P
- e') S57P+R170I+S188P+A194P
- f) Y167L+R170L
- f') Y167L+R170I
- g) Y167I+R170L
- 15 g') Y167I+R170I
- h) N76D+R170L+N218S
- h') N76D+R170I+N218S
- i) S57P+N76D+R170L+N218S
- i') S57P+N76D+R170I+N218S
- 20 j) N76D+R170L+N218S+M222A
- j') N76D+R170I+N218S+M222S
- j'') N76D+R170L+N218S+M222A
- j''') N76D+R170L+N218S+M222S
- k) S57P+R170I+S188P+A194P+N218S
- 25 k') S57P+R170I+S188P+A194P+N218S
- l) *36D+N76D+H120D+R170L+G195E+K235L
- l') *36D+N76D+H120D+R170I+G195E+K235L
- m) N76D+H120D+R170L+G195E+K235L
- m') N76D+H120D+R170I+G195E+K235L
- 30 n) *36D+G97N+V104Y+H120D+R170L+A194P+G195E+K235L
- n') *36D+G97N+V104Y+H120D+R170I+A194P+G195E+K235L
- o) S57P+R170L+Q206E
- o') S57P+R170I+Q206E
- p) R170L+Q206E
- 35 p') R170I+Q206E
- q) Y167I+R170L+Q206E
- q') Y167I+R170I+Q206E
- r) Y167F+R170L

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- x') Y167F+R170I
- t) Y167I+R170L+A194P
- t') Y167I+R170I+A194P
- u) Y167I+R170L+N218S
- 5 u) Y167I+R170I+N218S
- v) Y167I+R170L+A194P+N218S
- v') Y167I+R170I+A194P+N218S
- x) R170L+P131V
- x') R170I+P131V
- 10 y) *36D+Y167I+R170L
- y') *36D+Y167I+R170I
- z) Y167I+Y171I
- aa) Y167V+R170L
- aa') Y167V+R170I

15

DETERGENT COMPOSITIONS COMPRISING THE MUTANT ENZYMES

The present invention also comprises the use of the mutant enzymes of the invention in cleaning and detergent compositions and such compositions comprising the 20 mutant subtilisin enzymes. Such cleaning and detergent compositions can in principle have any physical form, but the subtilase variants are preferably incorporated in liquid detergent compositions or in detergent compositions in the form of bars, tablets, sticks and the like for direct 25 application, wherein they exhibit improved enzyme stability.

Among the detergent compositions of the present invention are first of all liquid detergents, especially aqueous liquid detergents having for example a homogeneous physical character, e.g. they can consist of a micellar 30 solution of surfactants in a continuous aqueous phase, so-called isotropic liquids.

Alternatively and preferably, they can have a heterogeneous physical phase and they can be structured, for example they can consist of a dispersion of lamellar 35 droplets in a continuous aqueous phase, for example comprising a deflocculating polymer having a hydrophilic backbone and at least one hydrophobic side chain, as described in EP-A-346 995 (Unilever) (incorporated herein by

reference). These latter liquids are heterogeneous and may contain suspended solid particles such as particles of builder materials e.g. of the kinds mentioned below.

Preferably, the liquid cleaning and detergent compositions of the invention should have a high electrolyte concentration, such as described in EP-A-328 177, EP-A-359 308, EP-A-328 176, EP-A-346 995 (all Unilever).

Such compositions comprise in addition to any one or more of the subtilisin enzyme variants in accordance to any of the preceding aspects of the invention alone or in combination any of the usual components included in such compositions which are well-known to the person skilled in the art.

Such components comprise builders, such as phosphate or zeolite builders, surfactants, such as anionic, cationic, non-ionic or zwitterionic type surfactants, polymers, such as acrylic or equivalent polymers, bleach systems, such as perborate- or amino-containing bleach precursors or activators, structurants, such as silicate structurants, alkali or acid to adjust pH, humectants, and/or neutral inorganic salts.

Furthermore, a number of optional ingredients are normally present in the compositions of the invention, such as:

- 25 A. Optional Cosurfactants
- B. Tartrate Succinate Builder
- C. Neutralization System
- D. Suds Suppressor
- E. Other Enzymes
- 30 F. Other Optional Components

The weight ratio of synthetic anionic surfactant to ethoxylated nonionic surfactant is from 1:1 to 5:1. The compositions have a pH in a 10% by weight solution in water at 20°C of from 7.0 to 9.0, a Critical Micelle Concentration of less than or equal to 200 ppm, and an air/water Interfacial Tension at the Critical Micelle Concentration of less than or equal to 32 dynes/cm at 35°C in distilled water. The compositions are preferably clear, homogeneous and phase

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stable, and have good cleaning performance and enzyme stability.

VARIOUS COMPONENTS:

5 1. Synthetic Anionic Surfactant

The liquid detergent compositions of the present invention contain from about 10% to about 50%, preferably from about 15% to about 50%, more preferably from about 20% to 40%, and most preferably from 20% to about 30%, by weight of a 10 natural or synthetic anionic surfactant. Suitable natural or synthetic anionic surfactants e.g. soaps and such as disclosed in US-A-4 285 841, and in US-A-3 929 678.

Useful anionic surfactants include the water-soluble salts, particularly the alkali metal, ammonium and 15 alkylolammonium (e.g., monoethanolammonium or triethanol-ammonium) salts, of organic sulfuric reaction products having in their molecular structure an alkyl group containing from about 10 to about 20 carbon atoms and a sulphonic acid or sulphuric acid ester group. (Included in 20 the term "alkyl" is the alkyl portion of aryl groups.) Examples of this group of synthetic surfactants are the alkyl sulfates, especially those obtained by sulfating the higher alcohols (C₈-C₁₈ carbon atoms) such as those produced by reducing the glycerides of tallow or coconut oil; and the 25 alkylbenzene sulfonates in which the alkyl group contains from about 9 to about 15 carbon atoms, in straight chain or branched chain configuration, e.g., those of the type described in U.S. Patents 2 220 099 and 2 477 383. Especially valuable are linear straight chain alkylbenzene 30 sulfonates in which the average number of carbon atoms in the alkyl group is from about 11 to 14.

Other anionic surfactants herein are the water-soluble salts of: paraffin sulfonates containing from 8 to about 24 (preferably about 12 to 18) carbon atoms; alkyl 35 glyceryl ether sulfonates, especially those ethers of C₈-C₁₈ alcohols (e.g., those derived from tallow and coconut oil); alkyl phenol ethylene oxide ether sulfates containing from 1 to about 4 units of ethylene oxide per molecule and from 8

to 12 carbon atoms in the alkyl group; and alkyl ethylene oxide ether sulfates containing 1 to 4 units of ethylene oxide per molecule and from 10 to 20 carbon atoms in the alkyl group.

5 Other useful anionic surfactants include the water-soluble salts of esters of α -sulfonated fatty acids containing from 6 to 20 carbon atoms in the fatty acid group and from 1 to 10 carbon atoms in the ester group; water-soluble salts of 2-acyloxy-alkane-1-sulphonic acids

10 containing from 2 to 9 carbon atoms in the acyl group and from 9 to 23 carbon atoms in the alkane moiety; water-soluble salts of olefin sulfonates containing from 12 to 24 carbon atoms; and β -alkyloxy alkane sulfonates containing from 1 to 3 carbon atoms in the alkyl group and from 8 to 20

15 carbon atoms in the alkane moiety.

Preferred anionic surfactants are the C_{10} - C_{12} alkyl sulfates and alkyl ethoxy sulfates containing an average of up to 4 ethylene oxide units per mole of alkyl sulfate, C_{11} - C_{13} linear alkyl benzene sulfonates, and mixtures thereof.

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2. Ethoxylated Nonionic Surfactant

A second optional ingredient is from 2% to 14% preferably from 2% to 8%, most preferably from 3% to 5% by weight, of an ethoxylated nonionic surfactant. The weight 25 ratio of synthetic anionic surfactant (on an acid basis) to nonionic surfactant is from 1:1 to 5:1 preferably from 2:1 to 5:1, most preferably from 3:1 to 4:1. This is to ensure the formation and adsorption of sufficient hardness surfactants at the air/water interface to provide good 30 greasy/oily soil removal.

The ethoxylated nonionic surfactant is of the formula $R^1(OC_2H_4)_nOH$, wherein R^1 is a C_{10} - C_{12} alkyl group or a C_{10} - C_{12} alkyl phenyl group, n is from 3 to 9, and said nonionic surfactant has an HLB (Hydrophilic-Lipophilic 35 Balance) of from 6 to 14, preferably from 10 to 13. These surfactants are more fully described in US-A-4 285 841, and US-A-4 284 532. Particularly preferred are condensation products of C_{11} - C_{13} alcohols with from 3 to 8 moles of

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ethylene oxide per mole of alcohol, e.g., C₁₂-C₁₈ alcohol condensed with about 6.5 moles of ethylene oxide per mole of alcohol. Other nonionic surfactants to be mentioned are APG, EGE and glucamide surfactants.

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3. Detergency Builder

Among the usual detergent ingredients which may be present in usual amounts in the detergent compositions of this invention are the following: The compositions may be 10 built or unbuilt, and may be of the zero-P type (i.e. not containing any phosphorus containing builders). Thus, the composition may contain in the aggregate for example from 1-50%, e.g. at least about 5% and often up to about 35-40% by weight, of one or more organic and/or inorganic builders. 15 Typical examples of builders include those already mentioned above, and more broadly include alkali metal ortho, pyro, and tripoly-phosphates, alkali metal carbonates, either alone or in admixture with calcite, alkali metal citrates, alkali metal nitrilotriacetates, carboxymethyloxysuccinates, 20 zeolites, polyacetalcarboxylates, and so on.

More specifically, the compositions herein contain from 5% to 20%, preferably from 10% to 15%, by weight of a detergency builder which can be a fatty acid containing from 10 to 18 carbon atoms and/or a polycarboxylate, zeolite, 25 polyphoshonate and/or polyphosphate a builder. Preferred are from 0 to 10% (more preferably from 3% to 10%) by weight of saturated fatty acids containing from 12 to 14 carbon atoms, along with from 0 to 10%, more preferably from 2% to 8%, most preferably from 2% to 5%, by weight of a 30 polycarboxylate builder, most preferably citric acid, in a weight ratio of from 1:1 to 3:1.

Since the proteolytic enzymes herein appear to provide optimum storage stability benefits versus other enzymes when the builder to water hardness ratio is close to 35 one, the compositions preferably contain sufficient builder to sequester from 2 to 10, preferably from 3 to 8, grains per gallon of hardness.

Suitable saturated fatty acids can be obtained

from natural sources such as plant or animal esters (e.g., palm kernel oil, palm oil and coconut oil) or synthetically prepared (e.g., via the oxidation of petroleum or by hydrogenation of carbon monoxide via the Fisher-Tropsch process). Examples of suitable saturated fatty acids for use in the compositions of this invention include capric, lauric, myristic, coconut and palm kernel fatty acid. Preferred are saturated coconut fatty acids; from 5:1 to 1:1 (preferably about 3:1) weight ratio mixtures of lauric and 10 myristic acid; mixtures of the above with minor amounts (e.g., 1%-30% of total fatty acid) of oleic acid; and palm kernel fatty acid.

The compositions herein preferably also contain the polycarboxylate, polyphosphonate and polyphosphate builders described in US-A-4 284 532. Water-soluble polycarboxylate builders, particularly citrates, are preferred of this group. Suitable polycarboxylate builders include the various aminopolycarboxylates, cycloalkane polycarboxylates, ether polycarboxylates, alkyl 20 polycarboxylates, epoxy polycarboxylates, tetrahydrofuran polycarboxylates, benzene polycarboxylates, and polyacetal polycarboxylates.

Examples of such polycarboxylate builders are sodium and potassium ethylenediaminetetraacetate; sodium and 25 potassium nitrilotriacetate; the water-soluble salts of phytic acid, e.g., sodium and potassium phytates, disclosed in US-A-1 739 942, the polycarboxylate materials described in US-A-3 364 103; and the water-soluble salts of polycarboxylate polymers and copolymers described in US-A-3 308 067.

Other useful detergency builders include the water-soluble salts of polymeric aliphatic polycarboxylic acids having the following structural and physical characteristics: (a) a minimum molecular weight of about 350 35 calculated as to the acid form; (b) an equivalent weight of 50 to 80 calculated as to acid form; (3) at least 45 mole percent of the monomeric species having at least two carboxyl radicals separated from each other by not more than

two carbon atoms: (d) the site of attachment of the polymer chain of any carboxyl-containing radical being separated by not more than three carbon atoms along the polymer chain from the site of attachment of the next carboxyl-containing radical. Specific examples of such builders are the polymers and copolymers of itaconic acid, aconitic acid, maleic acid, mesaconic acid, fumaric acid, methylene malonic acid, and citraconic acid.

Other suitable polycarboxylate builders include the water-soluble salts, especially the sodium and potassium salts, of mallic acid, citric acid, pyromellitic acid, benzene pentacarboxylic acid, oxydiacetic acid, carboxymethyloxysuccinic acid, carboxymethyloxymalonic acid, cis-cyclohexanehexacarboxylic acid, cis-cyclopentane-tetracarboxylic acid and oxydisuccinic acid.

Other polycarboxylates are the polyacetal carboxylates described in US-A-4 144 226, and US-A-4 146 495.

Other detergency builders include the zeolites, such as the aluminosilicate ion exchange material described in US-A-4 405 483.

Other preferred builders are those of the general formula R-CH(COOH)CH₂(COOH), i.e. derivatives of succinic acid, wherein R is C₁₂-C₂₂ alkyl or alkenyl, preferably C₁₂-C₁₆, or wherein R may be substituted with hydroxyl, sulfo, sulfoxy or sulfone substituents. These succinate builders are preferably used in the form of their water soluble salts, including the sodium, potassium and alkanolammonium salts. Specific examples of succinate builders include: lauryl succinate, myristyl succinate, palmityl succinate, 2-dodecenyl succinate, and the like.

4. Proteolytic Enzyme

The enzymes of the invention can be used in well-known standard amounts in detergent compositions. The amounts may range very widely, e.g. about 0.0002-0.1, e.g. about 0.005-0.05, Anson units per gram of the detergent.

composition. Expressed in alternative units, the protease can be included in the compositions in amounts in the order of from about 0.1 to 100 GU/mg (e.g. 1-50, especially 5-20 GU/mg) of the detergent formulation, or any amount in a wide range centering at about 0.01-4, e.g. 0.1-0.4 KNPU per g of detergent formulation.

It may for example be suitable to use the present enzymes at the rate of about 0.25 mg of enzyme protein per litre of wash liquor, corresponding to an enzyme activity of 10 the order of 0.08 KNPU per litre. Corresponding detergent formulations can contain the enzymes in for example an amount of the order of 0.1-0.4 KNPU/g.

Expressed differently the compositions of the present invention contain from about 0.01% to about 5%, 15 preferably from about 0.1% to about 2%, by weight of the proteolytic enzymes of the invention.

The described proteolytic enzyme is preferably included in an amount sufficient to provide an activity of from 0.05 to about 1.0, more preferably from about 0.1 to 20 0.75, most preferably from about 0.125 to about 0.5, mg of active enzyme per gram of composition.

5. Enzyme Stabilization System

The liquid detergents according to the present invention may comprise an enzyme stabilization system, 25 comprising calcium ion, boric acid, propylene glycol and/or short chain carboxylic acids. The enzyme stabilization system comprises from about 0.5% to about 15% by weight of the composition.

30 The composition preferably contains from about 0.01 to about 50, preferably from about 0.1 to about 30, more preferably from about 1 to 20 millimoles of calcium ion per liter. The level of calcium ion should be selected so that there is always some minimum level available for the 35 enzyme, after allowing for complexation with builders etc. in the composition. Any water-soluble calcium salt can be used as the source of calcium ion, including calcium chloride, calcium formate, and calcium acetate. A small

amount of calcium ion, generally from about 0.05 to 0.4 millimoles per liter, is often also present in the composition due to calcium in the enzyme slurry and formula water. From about 0.03% to about 0.6% of calcium formate is 5 preferred.

A second preferred enzyme stabilizer is polyols containing only carbon, hydrogen and oxygen atoms. They preferably contain from 2 to 6 carbon atoms and from 2 to 6 hydroxy groups. Examples include propylene glycol 10 (especially 1,2-propanediol, which is preferred), ethylene glycol, glycerol, sorbitol, mannitol, and glucose. The polyol generally represents from about 0.5% to 15%, preferably from about 1.5% to about 8%, by weight of the composition. Preferably, the weight ratio of polyol to any 15 boric acid added is at least 1, more preferably at least 1.3.

The compositions preferably also contain the water-soluble, short chain carboxylates described in US-A-4 318 818. The formates are preferred and can be used at 20 levels of from about 0.05% to about 5%, preferably from about 0.2% to about 2%, most preferably from 0.4% to 1.5%, by weight of the composition. Sodium formate is preferred.

The compositions herein also optionally contain from about 0.25% to about 5%, most preferably from about 25 0.5% to about 3%, by weight of boric acid. The boric acid may be, but is preferably not, formed by a compound capable of forming boric acid in the composition. Boric acid is preferred, although other compounds such as boric oxide, borax and other alkali metal borates (e.g., sodium ortho-, meta- and pyroborate, and sodium pentaborate) are suitable. (Substituted boric acids (e.g., phenylboronic acid, butane boronic acid, and p-bromo phenylboronic acid) can also be 30 used in place of boric acid.

35 6. Water

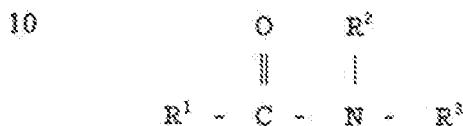
The liquid detergent compositions of the present invention may be aqueous liquids or non-aqueous liquids. When they are aqueous liquids, they contain from about 15% to

about 60%, preferably from about 25% to about 45%, by weight of water.

5 FURTHER OPTIONAL COMPONENTS

A. Optional Cosurfactants

Optional cosurfactants for use with the above ethoxylated nonionic surfactants include amides of the formula



wherein R¹ is an alkyl, hydroxyalkyl or alkenyl radical containing from 8 to 20 carbon atoms, and R² and R³ are selected from the group consisting of hydrogen, methyl, ethyl, propyl, isopropyl, 2-hydroxyethyl, 2-hydroxypropyl, 3-hydroxypropyl, and said radicals additionally containing up to 5 ethylene oxide units, provided at least one of R² and R³ contains a hydroxyl group.

Preferred amides are the C₈-C₂₀ fatty acid alkylol amides in which each alkylol group contains from 1 to 3 carbon atoms, and additionally can contain up to 2 ethylene oxide units. Particularly preferred are the C₁₂-C₁₆ fatty acid monoethanol and diethanol amides.

If used, amides are preferably present at a level such that the above ethoxylated nonionic surfactant and amide surfactant is in a weight ratio of from 4:1 to 1:4, preferably from 3:1 to 1:3.

30 Preferred and optional cosurfactants, used at a level of from 0.15% to 1%, are the quaternary ammonium, amine and amine oxide surfactants described in US-A-4 507 219.

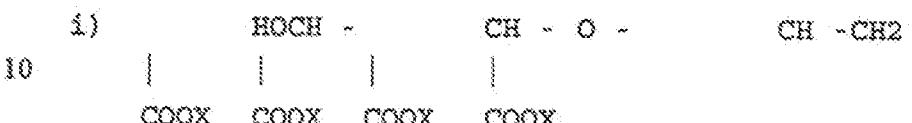
Of the above, the C₁₀-C₁₄ alkyl trimethylammonium salts are preferred, e.g., decyl trimethylammonium methylsulfate, lauryl trimethylammonium chloride, myristyl trimethylammonium bromide, and coconut trimethylammonium chloride and methylsulfate. From 0.2% to 0.8% of monoalkyl

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trimethylammonium chloride is preferred.

B. Tartrate Succinate Builder

The compositions herein preferably contain from 0 to about 10%, preferably from 0 to about 6%, by weight on an acid basis, of a tartrate succinate builder material selected from the group consisting of:



wherein X is a salt-forming cation;



wherein X is a salt-forming cation; and

20 iii) mixtures thereof.

The tartrate succinate compounds used herein are described in US-A-4 663 071.

25 C. Neutralization System

The present compositions can also optionally contain from about 0 to about 0.04 moles, preferably from about 0.01 to 0.035 moles, more preferably from about 0.015 to about 0.03 moles, per 100 grams of composition of an alkanolamine selected from the group consisting of monoethanolamine, diethanolamine, triethanolamine, and mixtures thereof. Low levels of the alkanolamines, particularly monoethanolamine, are preferred to enhance product stability, detergency performance, and odour. However, the amount of alkanolamine should be minimized for best chlorine bleach compatibility.

In addition, the compositions contain sodium ions, and preferably potassium ions, at a level sufficient to

neutralize the anionic species and provide the desired product pH.

D. Suds Suppressor

5 Another optional component for use in the liquid detergents herein is from 0 to about 1.5%, preferably from about 0.5% to about 1.0%, by weight of silicone based suds suppressor agent.

10 Silicones are widely known and taught for use as highly effective suds controlling agents. For example, US-A-3 455 839 relates to compositions and processes for defoaming aqueous solutions by incorporating therein small amounts of polydimethylsiloxane fluids.

15 Useful suds controlling silicones are mixtures of silicone and silanated silica as described, for instance, in German Patent Application DE-A-2 124 526.

20 Silicone defoamers and suds controlling agents have been successfully incorporated into granular detergent compositions by protecting them from detergent surfactants as in U.S. Patents 3 933 672 and 4 652 392.

A preferred silicone based suds suppressor for use herein is a suds suppressing amount of a suds controlling agent consisting essentially of:

25 (i) polydimethylsiloxane fluid having a viscosity of from about 20 cs. to about 1500 cs. at 25°C;

(ii) from about 5 to about 50 parts per 100 parts by weight of (i) of siloxane resin composed of $(CH_3)_2SiO_{1/2}$ units and SiO_2 units in a ratio of from $(CH_3)_2SiO_{1/2}$ units and to SiO_2 units of from about 0.6:1 to about 1.2:1; and

(iii) from about 1 to about 20 parts per 100 parts by weight of (i) of a solid silica gel.

35 By "suds suppressing amount" is meant that the formulator of the composition can select an amount of this

suds controlling agent that will control the suds to the extent desired. The amount of suds control will vary with the detergent surfactant selected. For example, with high sudsing surfactants, relatively more of the suds controlling agent is used to achieve the desired suds control than with low foaming surfactants.

E. Other Enzymes

The detergent compositions of the invention may also 10 contain further enzymes.

For example, lipase can usefully be added in the form of a (granular composition (alternatively a) solution or a slurry of lipolytic enzyme with carrier material (e.g. as in EP-A-258 068 (Novo Nordisk A/S)).

15 The added amount of lipase can be chosen within wide limits, for example 50 to 30,000 LU/g per gram of the surfactant system or of the detergent composition, e.g. often at least 100 LU/g, very usefully at least 500 LU/g, sometimes preferably above 1000, above 2000 LU/g or above 20 4000 LU/g or more, thus very often within the range of 50-4000 LU/g, and possibly within the range of 200-1000 LU/g. In this specification, lipase units are defined as they are in EP-A-258 068.

25 The lipolytic enzyme can be chosen among a wide range of lipases. In particular, the lipases described in for example the following patent specifications: EP-A-214 761 (Novo Nordisk A/S), EP-A-258 068, and especially lipases showing immunological cross reactivity with antisera raised against lipase from *Thermomyces lanuginosus* ATCC 22070, EP- 30 A-205 208 and EP-A-206 390, and especially lipases showing immunological cross-reactivity with antisera raised against lipase from *Chromobacter viscosum* var *lipolyticum* NRRL B-3673, or against lipase from *Alcaligenes* PL-679, ATCC 31371 and FERM-P 3783, also the lipases described in 35 specifications WO 87/00859 (Gist-Brocades) and EP-A-204 284 (Sapporo Breweries). Suitable, in particular, are for example the following commercially available lipase preparations: Lipolase® Novo Nordisk A/S, Amano lipases CE, P,

B, AP, M-AP, AML, and CES, and Maito lipases MY-30, OF, and PL, also Esterase[®] MM, Lipozym, SP225, SP285 (all Novo Nordisk), Saiken lipase, Enzeco lipase, Toyo Jozo lipase and Diosynth lipase (Trade Marks), Lumafast[®] (Genecor Inc.),
5 Lipomax[®] (Gist-Brocades N.V.), and lipases as described in WO-A-94/03578 (Unilever)

Amylase can for example be used when desired, in an amount in the range of about 1 to about 100 MU (maltose units) per gram of detergent composition (or 0.014-1.4, e.g. 10 0.07-0.7, KNU/g (Novo units)). Amylases suitable are for example Termamyl[®] and BAN (Novo Nordisk A/S). Cellulase can for example be used when desired, in an amount in the range of about 0.3 to about 35 CEVU units per gram of the detergent composition. Suitable cellulases are for example 15 Celluzyme[®] and Carezyme[®] (Novo Nordisk A/S).

Other enzymes contemplated to be used in the present invention are oxidases and peroxidases.

F. Other Optional Components

20 Other optional components for use in the liquid detergents herein include soil removal agents, soil release polymers, antiredeposition agents such as tetraethylene pentamine ethoxylate (from about 0.5% to 3%, preferably from about 1% to about 3%, by weight), suds regulants, 25 hydrotropes such as sodium cumene sulfonate, opacifiers, antioxidants, bactericides, dyes, perfumes, and brighteners known in the art. Such optional components generally represent less than about 15%, preferably from about 0.5% to 10%, more preferably from about 1% to about 10%, by weight 30 of the composition.

The compositions may contain from 0% to about 8%, preferably from 0% to about 5%, by weight of a C₁₂-C₁₄ alkenyl succinic acid or salt thereof. These materials are of the general formula R-CH(COOX)CH₂(COOX), wherein R is a C₁₂-C₁₄ alkenyl group and each X is H or a suitable cation, such as sodium, potassium, ammonium or alkanolammonium (e.g., mono-, di-, or tri-ethanolammonium). Specific examples are 2-dodecenyl succinate (preferred) and 2-tetradecenyl

succinate.

The compositions herein optionally contain from about 0.1% to about 1%, preferably from about 0.2% to about 0.6%, by weight of water-soluble salts of ethylenediamine

5 tetramethylenephosphonic acid, diethylenetriamine pentamethylenephosphonic acid, ethylenediamine tetraacetic acid (preferred), or diethylenetriamine pentaacetic acid (most preferred) to enhance cleaning performance when pretreating fabrics.

10 Furthermore, the detergent compositions may contain from 1-35% of a bleaching agent or a bleach precursor or a system comprising bleaching agent and/or precursor with activator therefor.

15 Further optional ingredients are lather boosters, anti-corrosion agents, soil-suspending agents, sequestering agents, anti-soil redeposition agents, and so on.

The compositions herein preferably contain up to about 10% of ethanol.

20 G. Other Properties

The instant composition usually has a pH, in a 10% by weight solution in water at 20°C, of from about 7.0 to 9.0, preferably from about 8.0 to about 8.5.

The instant compositions also may have a Critical 25 Micelle Concentration (CMC) of less than or equal to 200 parts per million (ppm), and an air/water Interfacial Tension above the CMC of less than or equal to 32, preferably less than or equal to about 30, dynes per centimetre at 35°C in distilled water. These measurements are 30 described in "Measurement of Interfacial Tension and Surface Tension - General Review for Practical Man" C. Weser, GIT Fachzeitschrift für das Laboratorium, 24 (1980) 642-648 and 734-742, FIT Verlag Ernst Giebeler, Darmstadt, and "Interfacial Phenomena - Equilibrium and Dynamic Effects", 35 C. A. Miller and P. Neogi, Chapter 1, pp. 29-36 (1985), Marcel Dekker, Inc. New York.

The compositions of the invention can be used for the washing of textile materials, especially, but without

limitation cotton and polyester based textiles and mixtures thereof. For example washing processes carried out at temperatures of about 60-65°C or lower, e.g. about 30-35°C or lower, are particularly suitable. It can be very suitable
5 to use the compositions at a rate sufficient to provide about e.g. 0.4-0.8 g/l of surfactant in the wash liquor, although it is of course possible to use lower or higher concentrations, if desired. Without limitation it can for example be stated that a use-rate from about 1 to 10 g/l,
10 e.g. from about 3-6 g/l, of the detergent formulation is suitable for use in the case when the formulations are substantially as in the Examples.

In this aspect the invention is especially related to:

15 a) A detergent composition formulated as an aqueous detergent liquid comprising anionic surfactant, nonionic surfactant, humectant, organic acid, caustic alkali, with a pH adjusted to a value between 9 and 10.

20 b) A detergent composition formulated as a non-aqueous detergent liquid comprising a liquid nonionic surfactant consisting essentially of linear alkoxylated primary alcohol, triacetin, sodium tripophosphate, caustic alkali, perborate monohydrate bleach precursor, and tertiary amine
25 bleach activator, with a pH adjusted to a value between about 9 and 10.

c) An enzymatic liquid detergent composition formulated to give a wash liquor pH of 9 or less when used at a rate
30 corresponding to 0.4-0.8 g/l surfactant.

d) An enzymatic liquid detergent composition formulated to give a wash liquor pH of 8.5 or more when used at a rate corresponding to 0.4-0.8 g/l surfactant.

35 e) An enzymatic liquid detergent composition formulated to give a wash liquor ionic strength of 0.03 or less, e.g. 0.02 or less, when used at a rate corresponding to 0.4-0.8

g/l surfactant.

5 f) An enzymatic liquid detergent composition formulated to give a wash liquor ionic strength of 0.01 or more, e.g. 0.02 or more, when used at a rate corresponding to 0.4-0.8 g/l surfactant.

10 It was found that the subtilase variants of the present invention can also be usefully incorporated in detergent composition in the form of bars, tablets, sticks and the like for direct application to fabrics, hard 15 surfaces or any other surface. In particular, they can be incorporated into soap or soap/synthetic compositions in bar form, wherein they exhibit a remarkable enzyme stability.

16 Detergent composition in the form of bars, tablets, sticks and the like for direct application, are for example described in South African Patent 93/7274, incorporated herein by reference. Accordingly, the preferred bars in accordance with this invention comprise, in addition to the subtilase variant:

20 i) 25 to 80%, most preferably 25 to 70%, by weight of detergent active which is soap or a mixture of soap and synthetic detergent active, reckoned as anhydrous;

25 ii) 0 to 50% and, most preferably, 10 to 30% by weight of water;

iii) 0 to 35% and, most preferably, 0.1 to 30% by weight filler.

30 In general, the amount of subtilase variant to be included in such compositions of the invention is such that it corresponds with a proteolytic activity of 0.1 to 100 GU/mg based on the composition, preferably 0.5 to 20GU/mg, most preferably 1.0 to 10GU/mg, where GU/mg is glycine unit per milligram.

35 METHOD FOR PRODUCING MUTATIONS IN SUBTILASE GENES

Many methods for introducing mutations into genes are well known in the art. After a brief discussion of cloning subtilase genes, methods for generating mutations in both

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random sites, and specific sites, within the subtilase gene will be discussed.

CLONING A SUBTILASE GENE

5 The gene encoding a subtilase may be cloned from any of the organisms indicated in Table I, especially gram-positive bacteria or fungus, by various methods, well known in the art. First a genomic, and/or cDNA library of DNA must be constructed using chromosomal DNA or messenger RNA from 10 the organism that produces the subtilase to be studied. Then, if the amino-acid sequence of the subtilase is known, homologous, labelled oligonucleotide probes may be synthesized and used to identify subtilisin-encoding clones from a genomic library of bacterial DNA, or from a cDNA library. 15 Alternatively, a labelled oligonucleotide probe containing sequences homologous to subtilase from another strain of bacteria or organism could be used as a probe to identify subtilase-encoding clones, using hybridization and washing conditions of lower stringency.

20 Yet another method for identifying subtilase-producing clones would involve inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming protease-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto 25 agar containing a substrate for subtilase, such as skim milk. Those bacteria containing subtilase-bearing plasmid will produce colonies surrounded by a halo of clear agar, due to digestion of the skim milk by excreted subtilase.

30 GENERATION OF RANDOM MUTATIONS IN THE SUBTILASE GENE

Once the subtilase gene has been cloned into a suitable vector, such as a plasmid, several methods can be used to introduce random mutations into the gene.

35 One method would be to incorporate the cloned subtilase gene, as part of a retrievable vector, into a mutator strain of *Escherichia coli*.

Another method would involve generating a single stranded form of the subtilase gene, and then annealing the

fragment of DNA containing the subtilase gene with another DNA fragment such that a portion of the subtilase gene remained single stranded. This discrete, single stranded region could then be exposed to any of a number of mutagenizing agents, including, but not limited to, sodium bisulfite, hydroxylamine, nitrous acid, formic acid, or hydralazine. A specific example of this method for generating random mutations is described by Shortle and Nathans (1978, *Proc. Natl. Acad. Sci. U.S.A.*, 75 2170-2174). According to the Shortle and Nathans method, the plasmid bearing the subtilase gene would be nicked by a restriction enzyme that cleaves within the gene. This nick would be widened into a gap using the exonuclease action of DNA polymerase I. The resulting single-stranded gap could then be mutagenized using any one of the above mentioned mutagenizing agents.

Alternatively, the subtilisin gene from a *Bacillus* species including the natural promoter and other control sequences could be cloned into a plasmid vector containing replicons for both *E. coli* and *B. subtilis*, a selectable phenotypic marker and the M13 origin of replication for production of single-stranded plasmid DNA upon superinfection with helper phage IRI. Single-stranded plasmid DNA containing the cloned subtilisin gene is isolated and annealed with a DNA fragment containing vector sequences but not the coding region of subtilisin, resulting in a gapped duplex molecule. Mutations are introduced into the subtilisin gene either with sodium bisulfite, nitrous acid or formic acid or by replication in a mutator strain of *E. coli* as described above. Since sodium bisulfite reacts exclusively with cytosine in a single-stranded DNA, the mutations created with this mutagen are restricted only to the coding regions. Reaction time and bisulfite concentration are varied in different experiments such that from one to five mutations are created per subtilisin gene on average. Incubation of 10 µg of gapped duplex DNA in 4 M Na-bisulfite, pH. 6.0, for 9 minutes at 37°C in a reaction volume of 400 µl, deaminates about 1% of cytosines in the

single-stranded region. The coding region of mature subtilisin contains about 200 cytosines, depending on the DNA strand. Advantageously, the reaction time is varied from about 4 minutes (to produce a mutation frequency of about 5 one in 200) to about 20 minutes (about 5 in 200).

After mutagenesis the gapped molecules are treated *in vitro* with DNA polymerase I (Klenow fragment) to make fully double-stranded molecules and fix the mutations. Competent *E. coli* are then transformed with the mutagenized DNA to produce an amplified library of mutant subtilisins.

Amplified mutant libraries can also be made by growing the plasmid DNA in a Mut D strain of *E. coli* which increases the range of mutations due to its error prone DNA polymerase.

The mutagens nitrous acid and formic acid may also be used to produce mutant libraries. Because these chemicals are not as specific for single-stranded DNA as sodium bisulfite, the mutagenesis reactions are performed according to the following procedure. The coding portion of the subtilisin gene is cloned in M13 phage by standard methods and single stranded phage DNA prepared. The single-stranded DNA is then reacted with 1 M nitrous acid pH. 4.3 for 15-60 minutes at 23°C or 2.4 M formic acid for 1-5 minutes at 23°C. These ranges of reaction times produce a mutation frequency of from 1 in 1000 to 5 in 1000. After mutagenesis, a universal primer is annealed to the M13 DNA and duplex DNA is synthesized using the mutagenized single-stranded DNA as a template so that the coding portion of the subtilisin gene becomes fully double-stranded. At this point the coding region can be cut out of the M13 vector with restriction enzymes and ligated into an un-mutagenized expression vector so that mutations occur only in the restriction fragment.

(Myers et al., *Science* 239 242-257 (1985)).

GENERATION OF SITE DIRECTED MUTATIONS IN THE SUBTILASE GENE

Once the subtilase gene has been cloned, and desirable sites for mutation identified and the residue to substitute for the original ones have been decided, these mutations can be introduced using synthetic

oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the desired mutation sites; mutant nucleotides are inserted during oligonucleotide synthesis. In a preferred method, a single stranded gap of DNA, 5 bridging the subtilase gene, is created in a vector bearing the subtilase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in by DNA polymerase I (Klenow fragment) and the construct is 10 ligated using T4 ligase. A specific example of this method is described in Morinaga et al., (1984, *Biotechnology* 2 646-639). According to Morinaga et al., a fragment within the gene is removed using restriction endonuclease. The vector/gene, now containing a gap, is then denatured and 15 hybridized to a vector/gene which, instead of containing a gap, has been cleaved with another restriction endonuclease at a site outside the area involved in the gap. A single-stranded region of the gene is then available for hybridization with mutated oligonucleotides, the remaining gap is 20 filled in by the Klenow fragment of DNA polymerase I, the insertions are ligated with T4 DNA ligase, and, after one cycle of replication, a double-stranded plasmid bearing the desired mutation is produced. The Morinaga method obviates the additional manipulation of constructing new restriction 25 sites, and therefore facilitates the generation of mutations at multiple sites. U.S. Reissue Patent number 34,606 by Estell et al., issued May 10, 1994, is able to introduce oligonucleotides bearing multiple mutations by performing minor alterations of the cassette, however, an even greater 30 variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be introduced.

EXPRESSION OF SUBTILASE MUTANTS

35 According to the invention, a mutated subtilase gene produced by methods described above, or any alternative methods known in the art, can be expressed, in enzyme form, using an expression vector. An expression vector generally

falls under the definition of a cloning vector, since an expression vector usually includes the components of a typical cloning vector, namely, an element that permits autonomous replication of the vector in a microorganism 5 independent of the genome of the microorganism, and one or more phenotypic markers for selection purposes. An expression vector includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or 10 various activator genes. To permit the secretion of the expressed protein, nucleotides encoding a "signal sequence" may be inserted prior to the coding sequence of the gene. For expression under the direction of control sequences, a target gene to be treated according to the invention is 15 operably linked to the control sequences in the proper reading frame. Promoter sequences that can be incorporated into plasmid vectors, and which can support the transcription of the mutant subtilase gene, include but are not limited to the prokaryotic β -lactamase promoter (Villa- 20 Kamaroff, et al. (1978) Proc. Natl. Acad. Sci. U.S.A. 75 3727-3731) and the tac promoter (DeBoer, et al. (1983) Proc. Natl. Acad. Sci. U.S.A. 80 21-25). Further references can also be found in "Useful proteins from recombinant bacteria" in Scientific American (1980) 242 74-94.

25 According to one embodiment *B. subtilis* is transformed by an expression vector carrying the mutated DNA. If expression is to take place in a secreting microorganism such as *B. subtilis* a signal sequence may follow the translation initiation signal and precede the DNA sequence of interest. 30 The signal sequence acts to transport the expression product to the cell wall where it is cleaved from the product upon secretion. The term "control sequences" as defined above is intended to include a signal sequence, when it is present.

Other host systems known to the skilled person are 35 also contemplated for the expression and production of the protease variants of the invention. Such host systems comprise fungi, including filamentous fungi, plant, avian and mammalian cells as well as others.

50

MATERIALS AND METHODSStrains:

B. subtilis 309 and 147 are variants of *Bacillus lentinus*, deposited with the NCIB and accorded the accession numbers 5 NCIB 10309 and 10147, and described in US-A-3 723 250 incorporated by reference herein.

E. coli MC 1000 (M.J. Casadaban and S.N. Cohen (1980); *J. Mol. Biol.* 138 179-207), was made r',m' by conventional 10 methods and is also described in US Patent Application Serial No. 039,298.

Proteolytic Activity

In the context of this invention proteolytic activity 15 is expressed in Kilo NOVO Protease Units (KNPU). The activity is determined relatively to an enzyme standard (SAVINASE®), and the determination is based on the digestion of a dimethyl casein (DMC) solution by the proteolytic enzyme at standard conditions, i.e. 50°C, pH 8.3, 9 min. 20 reaction time, 3 min. measuring time. A folder AF 220/1 is available upon request to Novo Nordisk A/S, Denmark, which folder is hereby included by reference.

A GU is a Glycine Unit, defined as the proteolytic enzyme activity which, under standard conditions, during a 25 15-minutes' incubation at 40 deg C, with N-acetyl casein as substrate, produces an amount of NH₂-group equivalent to 1 μ mole of glycine.

Enzyme activity can also be measured using the PNA assay, according to reaction with the soluble substrate 30 succinyl-alanine-alanine-proline-phenyl-alanine-para-nitrophenol, which is described in the Journal of American Oil Chemists Society, Rothgeb, T.M., Goodlander, B.D., Garrison, P.H., and Smith, L.A., (1988).

EXAMPLES

35 For the generation of enzyme variants according to the invention the same materials and methods as described in i.a. WO-A-89/06279 (Novo Nordisk A/S), EP-A-130 756 (Genentech), EP-A-479 870 (Novo Nordisk A/S), EP-A-214 435

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(Henkel), WO-A- 87/04461 (Amgen), WO-A-87/05050 (Genex), EP application no. 87303761 (Genentech), EP-A-260 105 (Genencor), WO-A-88/06624 (Gist-Brocades NV), WO-A-88/07578 (Genentech), WO-A-88/08028 (Genex), WO-A-88/08033 (Amgen), 5 WO-A-88/08164 (Genex), Thomas et al. (1985) *Nature*, 318 375-376; Thomas et al. (1987) *J. Mol. Biol.*, 193, 803-813; Russel and Persht (1987) *Nature* 328 496-500. Other methods well established in the art may also be used.

10 EXAMPLE 1

Construction and Expression of Enzyme Variants:

A vector suited to a synthetic gene coding for subtilase 309 and its mutants was constructed. It is essentially a pUC19 plasmid [Yanish-Perron and Messing (1985) 15 Gene; 33 103-119], in which the multiple cloning site has been replaced by a linker containing the restriction sites used to separate five sub-fragments constituting the gene. The new linker was inserted into EcoRI - HindIII cut pUC19 thereby destroying these sites. The details of this 20 construction are described in WO 92/19729 on pages 25-26 and in figure 1 (sheets 1/7-7/7) thereof, the content of which is hereby included by reference.

Each subfragment was made from 6 to 12 oligonucleotides. The oligonucleotides were synthesised on an 25 automatic DNA synthesizer using phosphoramidite chemistry on a controlled glass support [Beaucage and Carruthers (1981); *Tetrahedron Letters* 22 1859-1869].

The five subfragments were isolated on a 2% agarose gel and inserted into pSX191. The sequence was verified by 30 dideoxynucleotide sequencing. Fragments A-E were isolated and ligated together with KpnI-BamHI cut pSX191. The ligation mixtures were used to transform competent *E. coli* MC1000 r^-, m^+ selecting for ampicillin resistance. The 858 bp KpnI-BamHI fragment that constitutes the part of the 35 subtilisin 309 gene coding for the mature part of the enzyme was then used to replace the wild type gene on pSX212 giving rise to pSX222, which was then transformed into a competent *B. subtilis* strain. After fermentation of the transformed

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strain and purification of the enzyme it was shown that the product was indistinguishable from the wild type product.

Protease variants derived from the synthetic gene are made by using oligonucleotides with altered sequence at the 5 place(s) where mutation is wanted (e.g. with sequences as given below) and mixing them with the rest of the oligonucleotides appropriate to the synthetic gene. Assembly of the variant gene is carried out with the variant materials in a manner otherwise analogous to that described above.

10 Further information on synthetic genes generally is available in Agarval et al (1970); *Nature*; 227 27-34.

A KpnI site was introduced into the beginning of the subtilase 309 synthetic gene encoding the mature part of the enzyme. The method used is called oligonucleotide directed 15 double-strand break repair mutagenesis and is described by Mandecki (1986) *Proc. Nat. Acad. Sci. USA* 83 7177-7181.

pSX172 is opened with NcoI at the beginning of the mature part of the subtilase 309 gene and is mixed with the oligonucleotide NOR 789 (see WO-A-92/19729), heated to 100°C, 20 cooled to 0°C, and transformed into *E. coli*. After retransformation, the recombinants can be screened by colony hybridisation using 32-P-labelled NOR 789. The recombinants that turned out to be positive during the screening had the KpnI site introduced right in front of NcoI by changing two 25 bases without changing the amino acid sequence. pSX172 is described in EP Patent Publication No. 405 901. The KpnI site so created is inserted into pSX120 on a 400-bp Pvul-I-NheI fragment, giving rise to pSX212. pSX120 is also described in EP-A-405 901.

30 The synthetic gene is inserted between KpnI and BamHI on pSX212, giving rise to pSX222.

Examples of mutations and corresponding sequences of oligonucleotides are as follows:

35 R170L (fragment D1)

5' - AATTCAGGTGCAGGCTCAATCAGCTATCCGGCGCTCTAT - 3'

5' - ||||| * |||||

5' - GTCCACGTCCGAGTTAGTCGATAGGCCGCGAGATAACGCTTG - 3'

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R170I (fragment B1)

5' - AATTCAAGGTGCAGGCTCAATCAGCTATCCGGCGATCTAT - 3'
5' - GTCCACGTCCGAGTTAGTCGATAGCCGCTAGATAACGCTTC - 3'

S57P (fragment B1)

10 5' - AGCTTTGTACCACGGAACCGCCGACTCAAGATGG - 3'
10 3' - AACATGGTCCCCCTGGCGCTGAGTTCTACCCCTAACCC - 5'

These oligonucleotides were combined with the rest of the
15 oligonucleotides from the synthetic gene that was not chan-
ged.

EXAMPLE 2Purification of Enzyme Variants:

This procedure relates to purification of a 10
20 litre scale fermentation of the Subtilisin 147 enzyme, the
Subtilisin 309 enzyme or mutants thereof.

Approximately 8 litres of fermentation broth were
centrifuged at 6000 rpm for 35 minutes in 1 litre beakers.
The supernatants were adjusted to pH 6.5 using 10% acetic
25 acid and filtered on Seitz Supra S100 filter plates.

The filtrates were concentrated to approximately
400 ml using an Amicon CH2A UF unit equipped with an Amicon
S1Y10 UF cartridge. The UF concentrate was centrifuged and
filtered prior to absorption at room temperature on a
30 Bacitracin affinity column at pH 7. The protease was eluted
from the Bacitracin column at room temperature using 25% 2-
propanol and 1 M sodium chloride in a buffer solution with
0.01 dimethylglutaric acid, 0.1 M boric acid and 0.002 M
calcium chloride adjusted to pH 7.

35 The fractions with protease activity from the Ba-
citracin purification step were combined and applied to a
750 ml Sephadex G25 column (5 cm dia.) equilibrated with a
buffer containing 0.01 dimethylglutaric acid, 0.2 M boric
acid and 0.002 M calcium chloride adjusted to pH 6.5.

40 Fractions with proteolytic activity from the

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Sephadex G25 column were combined and applied to a 150 ml CM Sepharose CL 6B cation exchange column (5 cm dia.) equilibrated with a buffer containing 0.01 M dimethylglutaric acid, 0.2 M boric acid, and 0.002 M calcium chloride adjusted to 5 pH 6.5.

The protease was eluted using a linear gradient of 0-0.1 M sodium chloride in 2 litres of the same buffer (0-0.2 M sodium chloride in case of Subtilisin 147).

10 In a final purification step protease containing fractions from the CM Sepharose column were combined and concentrated in an Amicon ultrafiltration cell equipped with a GR81PP membrane (from the Danish Sugar Factories Inc.).

15 By using the techniques of Example 1 for the construction and the above isolation procedure the following subtilisin 309 variants were produced and isolated:

- A: G159I
- B: S164I
- C: Y167I
- D: R170I
- 20 E: R170L
- F: R170M
- G: R170F
- H: G195P
- I: S57P+R170L
- 25 J: R170L+N218S
- K: S57P+R170L+N218S
- L: R170L+N218S+M222A
- M: S57P+R170L+S188P+A194P
- N: Y167I+R170L
- 30 O: S57P+R170L+Q206E
- P: R170L+Q206E
- Q: Y167I+R170L+Q206E
- R: Y167I+R170L+A194P
- S: Y167I+R170L+N218S
- 35 T: Y167I+R170L+A194P+N218S
- U: Y167I+Y171I
- V: R170G
- W: R170C

EXAMPLE 3

DETERGENT COMPOSITIONS COMPRISING ENZYME VARIANTSExample D1:

5 An (isotropic) aqueous detergent liquid according to an embodiment of the invention is formulated to contain:

	<u>Ingredient</u>	<u>%</u>
10	NaLAS	8.0
	Neodol 25-9	8.0
	AES 25-3S	14.0
	NaCitrato.2H2O	5.0
	Propylene Glycol	5.0
15	Sorbitol	4.5
	F-dye Tinopal	0.15
	UNPA-GX	
	Lytron 614	0.03
	Opacifier	
20	Kathon	0,0003
	Preservative	
	Acid Blue 80	0.00117
	Acid Violet 48	0.0033
	Savinase 16L	0.25
25	Lipolase 100L	0.70
	Fragrance	0.15
	Water	ad 100.0

The pH is adjusted to 7.1.

§

Table III.

Residual enzyme activity (in percentage of original activity) after storage at 37°C for Example D1 comprising the BLS309 variant S57P+R170L+N218S.

	Storage time (in days)	Wild-type	S57P+R170L+N218S
0		100	100
3		44	74
7		11	50
10		8	36
14		7	27

From Table III it is evident that the variant S57P+R170L+N218S exhibits a remarkably improved stability in this type of detergent. Moreover, the variant S57P+R170L+N218S possesses excellent compatibility towards lipase.

Table IV.

Residual lipase activity (in percentage of original activity) after storage at 37°C for Example D1 comprising the BLS309 variant S57P+R170L+N218S and Lipolase (TM).

	Storage time (in days)	Lipolase plus: Wild-type S57P+R170L+N218S
0	100	100
3	38	67
7	24	44
10	22	33
14	21	27

From the above Table IV it is apparent that, in addition to the stability of the protease, the compatibility of the protease towards Lipolase is also improved.

35 Example D2i

A non-aqueous detergent liquid according to an embodiment of the invention is formulated using 38.5% C13-C15 linear primary alcohol alkoxylated with 4.9 mol/mol ethylene oxide

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and 2.7 mol/mol propylene oxide, 5% triacetin, 30% sodium triphosphate, 4% soda ash, 15.5% sodium perborate monohydrate containing a minor proportion of oxoborate, 4% TAED, 0.25% EDTA of which 0.1% as phosphonic acid, Aerosil 5 0.6%, SCMC 1%, and 0.6% protease. The pH is adjusted to a value between 9 and 10, e.g. about 9.8.

Example D3:

Structured liquid detergents can for example 10 contain, in addition to a protease as described herein, 2-15% nonionic surfactant, 5-40% total surfactant, comprising nonionic and optionally anionic surfactant, 5-35% phosphate-containing or non-phosphate containing builder, 0.2-0.8% polymeric thickener, e.g. cross-linked acrylic 15 polymer with m.w. over 10⁶, at least 10% sodium silicate, e.g. as neutral waterglass, alkali (e.g. potassium-containing alkali) to adjust to desired pH, preferably in the range 9-10 or upwards, e.g. above pH 11, with a ratio sodium cation: silicate anion (as free silica) 20 (by weight) less than 0.7:1, and viscosity of 0.3-30 Pas (at 20°C and 20⁻¹).

Suitable examples contain about 5% nonionic surfactant C13-15 alcohol alkoxylated with about 5 EO groups per mole and with about 2.7 PO groups per mole, 15-23% 25 neutral waterglass with 3.5 weight ratio between silica and sodium oxide, 13-19% KOH, 8-23% STPP, 0-11% sodium carbonate, 0.5% Carbopol 941 (TM).

Protease may be incorporated at for example 0.5%.

Example D4:

(Decoupling polymer liquid)

Prichlene 6907	4.5
KOH	10
Ethoxylated Alcohol.7EO (Synperonic A7)	4.5
35 Ethoxylated Alcohol.3EO (Synperonic A3)	4.5
Zeolite 4A	15
Fluorescer Tinopal CBS-X	0.08
Narlex DCI	1

Citric acid	8.23
Antifoam silicone DB100	0.3
LAS acid	16.5
Perfume	0.5
5 Water to	100%

Table V.

Residual enzyme activity (in percentage of original activity) after storage at 37°C for Example D4 comprising 10 the R170L variant of BLS309.

Storage time (days)	R170L	Wild-type
0	100	100
2	98	73
15 4	96	66
10	94	46
33	87	8
81	78	2.1
101	71	0

20

From Table V it is evident that the R170L variant exhibits a remarkably improved stability in this type of detergent.

Example D5:

25 (Decoupling polymer liquid)	
Priolene 6907	4.5
KOH	10
Ethoxylated Alcohol.7EO (Syperonic A7)	4.5
Ethoxylated Alcohol.3EO (Syperonic A3)	4.5
30 Zeolite 4A	15
Fluorescer Tinopal CBS-X	0.08
Narlex DCI	1
Citric acid	8.23
Antifoam silicone DB100	0.3
35 LAS acid	16.5
Lipolase 100L	0.6
Perfume	0.5
Water to	100%

Table VI.

Residual enzyme activity (in percentage of original activity) after storage at 37°C for Example D5 comprising the BLS309 variant S57P+R170L+N218S.

5

Storage time (days)	Residual Protease Activity		Residual Lipase Activity	
	S57P+R170L+N218S	Wildtype	Wildtype	Wildtype
S57P+R170L+N218S				
0	100	100	100	100
2	-	75	41	94
5	97	50	15	76
8	87	30	7	71
15	91	20	12	78
28	100	18	12	70

From Table VI it is evident that the variant S57P+R170L+N218S exhibits a remarkably improved stability in this type of detergent. Moreover, the variant S57P+R170L+N218S possesses excellent compatibility towards lipase.

Example D6:

25 Soap bars were produced containing 49.7 wt.% 80/20 tallow /coconut soap, 49.0% water, 20% sodium citrate, 1.0% citric acid and 0.031% protease. After preparation of the soap bars they were stored at ambient temperature and after specific time intervals samples were taken and measured for protease activity. The stability data are given below in Table VII:

storage (days)	W.T.	R170L	R170L+N218S+S57P	R170L+Y167I
0	100	100	100	100
1	50	100	97	94
2	25	91	100	83
3		100	94	80

60

6		98	89	90
10	0	100	94	71
17		93	80	73
27		95	86	70

5

From the above Table VII it is evident that the subtilase variants R170L, R170L+N218S+S57P and R170L+Y167I exhibit a remarkably improved stability in this type of detergent.

10 Example D7:

Soap bars were produced containing 63.88% 80/20 tallow/coconut soap, 1% coconut fatty acid, 25.1% water, 10% sodium citrate and 0.021% protease. The laundry soap bars were stored at 37°C and after specific time intervals 15 samples were taken and measured for protease activity.

Stability data:

storage (days)	W.T.	R170L+N218S+S57P
0	100	100
10		90.1
14		81.5
18	10	
20	0	91.4
31		72.8
35		79
45		78

20 From the above Table VIII it is evident that the subtilase variant R170L+N218S+S57P exhibits a remarkably improved 25 stability in this type of detergent.

EXAMPLE 4

Wash performance of detergent compositions comprising enzyme variants.

5 The following examples provide results from a number of washing tests that were conducted under the conditions indicated.

10 Table IX: Experimental conditions for evaluation of Subtilisin 309 variants.

Detergent	Protease model detergent '95
Detergent dose	3 g/l
pH	9.5
15 Wash time	15 min.
Temperature	15°C
Water hardness	9°dH ~ 1.61 mM Ca ⁺⁺ /Mg ⁺⁺
Enzymes	Subtilisin 309 variants as listed below
20 Enzyme conc.	0; 0.1, 0.2, 0.3, 0.4, 0.5, 1.0, 2.0, 3.0 mg/l
Test method	Miniwash *(SOP DEF-SM-0026.01/01)*
Swatch/volume	5 swatches (Ø 2.5 cm) / 50 ml
25 Test material	Grass on cotton (rinsed in water); *DF-9417718*

The above model detergent is a simple detergent formulation. The most characteristic features are that STP is used as builder and the content of anionic tenside (LAS) is quite high. Further the pH is adjusted to 9.5, which is low for a powder detergent.

30 The composition of the model detergent is as follows:

Table X

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25 % STP ($\text{Na}_3\text{P}_2\text{O}_{10}$)
 25 % Na_2SO_4
 10 % Na_2CO_3
 20 % LAS (Nansa SOS)
 5.5 % Nonionic (Dobanol 25-7)
 5 % $\text{Na}_2\text{Si}_2\text{O}_5$
 0.5 % Carboxymethylcellulose (CMC)
 9.5 % water

dose: 3 g/l

10 pH is adjusted to 9.5

Measurement of remmision (R) on the testmaterial has been done at 460 nm using an Elrepho 2000 photometer (without UV). The measured values have been fitted to the expression:

$$15 \quad \Delta R = \frac{a \cdot \Delta R_{\max} \cdot c}{\Delta R_{\max} + a \cdot c}$$

The improvement factor is calculated by use of the initial

$$\text{slope of the curve: } IF = \frac{a}{a_{ref.}}$$

ΔR is the wash effect of the enzyme in remmision units.

a is the initial slope of the fitted curve ($c=0$).

20 $a_{ref.}$ is the initial slope for the reference enzyme.

c is the enzyme concentration in mg/l

ΔE_{max} is the theoretical maximum wash effect of the enzyme in remmision units (c- μ m).

Table XI: Variants and improvement factors for Subtilisin 309.

Designation	Variant	IF
S003*	R170Y	2.8
S004	R170Y + G195E	2.6
S012*	R170Y + G195E + K251E	1.6
G	R170F	3.3
E	R170L	3.8
F	R170M	2.4
D	R170I	4.1
I	S57P + R170L	3.9
J	R170L + N218S	1.6
K	S57P + R170L + N218S	2.3
N	Y167I + R170L	6.2
P	R170L + Q206E	2.6
V	R170G	2.0
W	R170C	3.4
O	S57P + R170L + Q206E	2.9
Q	Y167I + R170L + Q206E	2.4

* Described in WO-A-91/00345

As it can be seen from Table XI, all the Subtilisin 309 variants of the invention exhibit an improvement in wash performance.

CLAIMS

1. A detergent composition comprising a subtilase variant wherein one or more amino acid residues situated in or in the vicinity of a hydrophobic domain of the parent subtilase have been substituted for an amino acid residue more hydrophobic than the original residue, said hydrophobic domain comprising the residues P129, P131, I165, Y167, Y171 of BLS309 (in BASBPN numbering), and the residues in the vicinity thereof comprises residues corresponding to the residues E136, G159, S164, R170, A194 and G195 of BLS309 (in BASBPN numbering), with the exception of the R170M, R170I and R170V variants of BASP92.
2. Composition according to claim 1, comprising a subtilase variant wherein the original amino acid residue has been substituted for a residue selected from the group comprising Val (V), Ile (I), Leu (L), Met (M), Phe (F), and Trp (W), preferably Val, Ile or Leu.
3. Composition according to claims 1 or 2, wherein the parent subtilase is chosen from the sub-group I-S1.
4. Composition according to claim 3, wherein the parent subtilase is chosen from the group comprising ABSS168, BASBPN, BSSDY, and BLSCAR.
5. Composition according to claims 1 or 2, wherein the parent subtilase is chosen from the sub-group I-S2.

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6. Composition according to claim 5, wherein the parent subtilase is chosen from the group comprising BLS147, BLS309, BAPB92 AND BYSYAB.

7. Composition according to claim 5, wherein the parent subtilase is TVTHER.

8. Composition according to any of claims 1 to 7, wherein said substitution(s) is/are combined with substitutions, insertions or deletions in any other position.

9. Composition according to claim 8, wherein said substitution(s) is/are combined with substitutions, insertions or deletions in any of the positions 36, 222, 218, 76.

10. Composition according to any of claims 1 to 9, being any of the following:

K136V, K136I, K136L, K136M, K136F,

S159V, S159I, S159L, S159M, S159F,

T164V, T164I, T164L, T164M, T164F,

K170V, K170I, K170L, K170M, K170F,

E195V, E195I, E195L, E195M, E195F,

Q136V, Q136I, Q136L, Q136M, Q136F,

T159V, T159I, T159L, T159M, T159F,

A164V, A164I, A164L, A164M, A164F,

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Y170V, Y170I, Y170L, Y170M, Y170F,
S194V, S194I, S194L, S194M, S194F,
E136V, E136I, E136L, E136M, E136F,
G159V, G159I, G159L, G159M, G159F,
G164V, G164I, G164L, G164M, G164F,
S164V, S164I, S164L, S164M, S164F,
R170V, R170I, R170L, R170M, R170F,
A194V, A194I, A194L, A194M, A194F,
P194V, P194I, P194L, P194M, P194F,
E195V, E195I, E195L, E195M, E195F,
G195V, G195I, G195L, G195M, G195F,

11. Composition according to any of claims 1 to 10, wherein said variant is combined with further substitutions, deletions and/or insertions in any one or more of the positions:

36, 57, 76, 218, 222, and 224.

12. Compositions according to claim 11, wherein said subtilase belongs to the I-82 sub-group and said further change is chosen from the group comprising *36D, S57P, N76D, N218S, M222S, M222A, and T224S.

13. Composition according to claim 12, wherein the variant is chosen from the group of variants comprising:

a) S57P+E170L

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- a') S57P+R170I
- b) R170L+N218S
- b') R170I+N218S
- c) S57P+R170L+N218S
- c') S57P+R170I+N218S
- c'') S57P+V104Y+R170L+N218S
- c''') S57P+V104Y+R170I+N218S
- d) R170L+N218S+M222A
- d') R170I+N218S+M222S
- d'') R170L+N218S+M222A
- d''') R170I+N218S+M222S
- e) S57P+R170L+S188P+A194P
- e') S57P+R170I+S188P+A194P
- f) Y167L+R170L
- f') Y167L+R170I
- g) Y167I+R170L
- g') Y167I+R170I
- h) N76D+R170L+N218S
- h') N76D+R170I+N218S
- i) S57P+N76D+R170L+N218S
- i') S57P+N76D+R170I+N218S
- j) N76D+R170L+N218S+M222A
- j') N76D+R170I+N218S+M222S

- j") N76D+R170L+N218S+M222A
- j'') N76D+R170L+N218S+M222S
- k) S57P+R170I+S188P+A194P+N218S
- k') S57P+R170I+S188P+A194P+N218S
- l) *36D+N76D+H120D+R170L+G195E+K235L
- l') *36D+N76D+H120D+R170I+G195E+K235L
- m) N76D+H120D+R170L+G195E+K235L
- m') N76D+H120D+R170I+G195E+K235L
- n) *36D+G97N+V104Y+H120D+R170L+A194P+G195E+K235L
- n') *36D+G97N+V104Y+H120D+R170I+A194P+G195E+K235L
- o) S57P+R170L+Q206E
- o') S57P+R170I+Q206E
- p) R170L+Q206E
- p') R170I+Q206E
- q) Y167I+R170L+Q206E
- q') Y167I+R170I+Q206E
- r) Y167F+R170L
- r') Y167F+R170I
- t) Y167I+R170L+A194P
- t') Y167I+R170I+A194P
- u) Y167I+R170L+N218S
- u) Y167I+R170I+N218S
- v) Y167I+R170L+A194P+N218S

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- v') Y167I+R170I+A194P+N218S
- x) R170I+P131V
- x') R170I+P131V
- y) *36D+Y167I+R170L
- y') *36D+Y167I+R170I
- z) Y167I+Y171I
- aa) Y167V+R170L
- aa') Y167V+R170I

14. Composition according to claim 1-13, wherein the subtilase variant is R170L or R170I and the composition is a liquid detergent and comprises a deflocculating polymer.

15. Composition according to claim 14, wherein the subtilase variant is S57P+V104Y+R170L+N218S or S57P+V104Y+R170I+N218S.

16. Composition according to claim 1-13, in shaped solid form.

Fig. 1.

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Fig.2.

